

Roles of Different T-Cell Subsets in Control of Herpes Simplex Virus Infection Determined by Using T-Cell-Deficient Mouse Models

ELANCHEZHIAN MANICKAN AND BARRY T. ROUSE*

*Department of Microbiology, College of Veterinary Medicine,
 University of Tennessee, Knoxville, Tennessee 37996-0845*

Received 23 June 1995/Accepted 14 September 1995

Herpes simplex virus infection of the scarified dermis results in infection of the nervous system and, subsequently, a cutaneous lesion in the innervated dermatome. We compared the pathogenesis of such zosteriform lesions in mice lacking or severely depleted of CD4⁺ or CD8⁺ T cells because of targeted gene disruption. Mice without CD4⁺ cells showed markedly increased susceptibility, whereas β 2 microglobulin knockout mice lacking CD8⁺ T cells were as resistant to challenge as were immunocompetent mice with the same genetic background. Our results demonstrate that CD4⁺ T cells are of primary importance in the control of herpes simplex virus infections of the skin and nervous system.

Herpes simplex virus (HSV) types 1 and 2 (HSV-1 and HSV-2, respectively) cause a wide spectrum of human infections. After an acute episode, the virus invariably ascends to the neural ganglia, where it remains latent indefinitely (10). During reactivation, the virus undergoes rounds of replication and migrates via the sensory nerve to the original site of acute infection, where it may cause recurrent lesions. The mouse zosteriform model resembles this pathogenesis except that the skin lesions result from spread from ganglia following primary cutaneous infection (12). We and others have used the mouse zosteriform model to assess vaccine effectiveness and to determine immunological mechanisms involved in antiviral immunity (4–6, 13). Such studies indicate that virus clearance from the cutaneous lesions depends largely on antibody and CD4⁺ T-cell function (4, 6, 7, 12). The dominant role of the latter cell type became evident in comparisons of the effectiveness of immune populations of CD4⁺ and CD8⁺ cells at preventing zosteriform lesions in nude mice or mice with severe combined immunodeficiency given adoptive transfers of individual cell types (4). In fact, the cytokine profiles induced by viral antigen

stimulation of the transferred population indicated that CD4⁺ T cells of the type 1 cytokine-producing phenotype are involved in viral clearance. In a model measuring the rapidity of viral clearance from a subcutaneous site, both CD4⁺ and CD8⁺ T cells were shown to function and both subsets appeared to operate by the release of gamma interferon (14).

To gain further insight into the cellular mechanisms which contribute to viral clearance and hence recovery from HSV infection, the susceptibility of mice lacking CD4⁺ (8) or CD8⁺ (2) T cells because of gene knockout were compared for susceptibility to HSV-induced zosterification.

Animals were infected on the shaved, scarified left flank with either a high dose (for both HSV-1.17 and HSV-2-MS, it was 10⁸ 50% tissue culture-infective doses [TCID₅₀]) or a low dose (for HSV-1, 10⁴ TCID₅₀; for HSV-2, 5.2 × 10³ TCID₅₀) of virus as described elsewhere (4, 5). Animals were observed daily for signs of lesion development and severity. Severity was judged subjectively in accordance with the following criteria: 1+, vesicle formation; 2+, local erosion and ulceration of the

TABLE 1. Susceptibility of mice genetically deficient in CD4⁺ T cells to HSV-1 and HSV-2 infections^a

Day post-challenge	CD4 -/-				CD4 +/-				C57.BL/6			
	HSV-1.17		HSV-2-MS		HSV-1.17		HSV-2-MS		HSV-1.17		HSV-2-MS	
	LD	HD	LD	HD	LD	HD	LD	HD	LD	HD	LD	HD
5	—	10 (1.0)	20 (1.0)	60 (1.0)	—	—	—	—	—	—	—	—
7	40 (2.0)	80 (2.5)	60 (3.7)	100 (3.8)	—	40 (2.0)	—	40 (2.0)	—	40 (2.0)	—	40 (2.0)
9	90 (3.1)	100 (3.7)	100 (4.0)	NS	—	100 (3.5)	—	80 (3.3)	—	80 (3.0)	—	80 (3.5)
11	100 (4.0)	NS	NS	NS	—	100 (4.0)	—	100 (4.0)	—	100 (4.0)	—	100 (4.0)
14	NS	NS	NS	NS	—	NS	—	NS	—	NS	—	NS

^a —, no lesion development; NS, no survivors. Anesthetized homozygous heterozygous, and C57.BL/6 mice were shaved, and the skin of the midflank was scarified. The animals were infected with low (LD) and high (HD) doses of HSV-1.17 or HSV-2-MS. For HSV-1.17, the low dose was 10⁴ TCID₅₀ and the high dose was 10⁸ TCID₅₀. For HSV-2-MS, the low dose was 5.2 × 10³ TCID₅₀ and the high dose was 10⁸ TCID₅₀. Shown are the percentages of animals that developed zoster lesions and the mean clinical severity (in parentheses). The data represent one of the three experiments conducted with 10 mice per group. CD4 -/- and CD4 +/- mice were kindly provided by Dan R. Littman, Howard Hughes Medical Institute, San Francisco, Calif. The mice were bred by Virginia Godfrey, Oak Ridge National Research Laboratory, Oak Ridge, Tenn. The offspring from heterozygous matings that were CD4 -/- were excluded.

* Corresponding author. Mailing address: Department of Microbiology, M409 Walters Life Sciences Building, Knoxville, TN 37996-0845. Phone: (615) 974-4026. Fax: (615) 974-4007.

TABLE 2. Susceptibility of $\beta 2$ microglobulin-deficient mice to HSV-1 and HSV-2 infections^a

Day post-challenge	$\beta 2M^{-/-}$				C57.BL/6			
	HSV-1.17		HSV-2-MS		HSV-1.17		HSV-2-MS	
	LD	HD	LD	HD	LD	HD	LD	HD
5	—	—	—	—	—	—	—	—
7	—	30 (2.0)	—	60 (1.7)	—	40 (2.0)	—	50 (2.0)
9	—	90 (3.0)	—	100 (3.4)	—	80 (3.0)	—	90 (3.0)
11	—	100 (4.0)	—	100 (4.0)	—	100 (4.0)	—	100 (4.0)
14	—	NS	—	NS	—	NS	—	NS

^a —, no lesion development; NS, no survivors; LD, low dose; HD, high dose. Homozygous and C57.BL/6 mice were anesthetized and skin of the midflank was depilated. A low or high-dose of HSV-1 or HSV-2 (as mentioned in the Table 1 footnote) was used to infect the mice. Shown are the percentages of mice that developed lesions; the values in parentheses are mean clinical scores. The data represent one of three experiments done with 10 mice per group. $\beta 2$ microglobulin $^{-/-}$ mice were obtained from R. Jaenisch, Howard Hughes Medical Institute. The mice which had been backcrossed five times were further backcrossed for five more generations with C57.BL/6 mice by Virginia Godfrey, Oak Ridge National Research Laboratory.

local lesion; 3+, mild-to-moderate ulceration; 4+, severe ulceration, hind limb paralysis, and encephalitis.

Mice vary markedly in susceptibility to HSV infection. Thus, the low dose of both HSV-1 and HSV-2 chosen for study was lethal to susceptible mouse strains, such as BALB/c. C57BL6 mice, however, were totally resistant to the low-dose challenge. However, C57BL6 mice were susceptible and developed zosteriform lesions and usually a lethal encephalomyelitis following infection with the high challenge dose of either HSV-1 or HSV-2 (Table 1). Mice virtually lacking CD8⁺ T cells because of targeted deletion of the $\beta 2$ microglobulin gene and backcrossed more than 10 times with strain C57BL6 retained the resistance pattern of C57BL6 mice, succumbing to a high-dose challenge but resisting a low-dose challenge (Table 2). On the other hand, mice lacking CD4⁺ T cells were highly susceptible to the low-dose challenge of both HSV-1 and HSV-2 (Table 1). In the experiments with CD4^{-/-} mice, heterozygous mice with the identical genetic background were also investigated. As is evident in Table 1, the heterozygous mice retained the high-resistance pattern of immunocompetent C57BL6 mice.

Taken together, the results confirm that immunity to cutaneous infection with HSV is essentially a property of CD4⁺ T cells. Moreover, if adequate clearance of the infection does not occur, the virus enters the nervous system and the animal usually succumbs to encephalitis. Indeed, we have observed only very few animals which developed zosteriform lesions and survived. Apparently, once the virus is within the nervous system and disseminates to the skin it usually also spreads to the brain and causes death. This result always occurs with HSV-2. Our data indicate that effective immunity to HSV can be attained without CD8⁺ T-cell function. Although $\beta 2$ microglobulin $^{-/-}$ mice do have very few CD8⁺ T cells (3), the ability to generate HSV-specific CD8⁺ cytotoxic T-lymphocyte responses is virtually absent (9). Accordingly, we suggest that the CD8⁺-mediated cytotoxic T-lymphocyte response, essen-

tial for immunity to several viruses, can be dispensed with for HSV. Indeed, in humans, the anti-HSV cytotoxic T-lymphocyte response is mediated largely by CD4⁺ T cells (11) and it seems that HSV expresses strategies that minimize or prevent cytotoxic T-lymphocyte recognition (1, 15).

In conclusion, the results of the present study with knockout mice confirm the primary importance of CD4⁺ T cells for immunity to HSV. The present study sheds no light on how CD4⁺ T cells subserve this function, although the work of others indicates that the elaboration of type 1 cytokines, particularly gamma interferon, may be the crucial function involved.

We thank Rudi Jaenisch and Dan Littman for the initial supply of knockout mice and Virginia Godfrey for their further breeding and backcrossing. We appreciate the assistance of Paula Keaton and Kim Cox.

This study was supported by NIH grant AI14981.

REFERENCES

1. Banks, T. A., and B. T. Rouse. 1992. On the immune evasive characteristics of herpesviruses. *Clin. Infect. Dis.* **14**:933-941.
2. Grusby, M. J., H. J. Archinross, R. Lee, R. S. Johnson, J. P. Spencer, M. Zijestra, R. Jaenisch, V. E. Papaioannou, and L. H. Glimcher. 1993. Mice lacking major histocompatibility complex class I and class II molecules. *Proc. Natl. Acad. Sci. USA* **90**:3913-3917.
3. Lamouse-Smith, E., V. K. Clements, and S. Ostrand-Rosenberg. 1993. $\beta 2M^{-/-}$ knockout mice contain low levels of CD8⁺ cytotoxic T-lymphocyte that mediate specific tumor rejection. *J. Immunol.* **151**:6283-6290.
4. Manickan, E., M. Francotte, N. Kuklin, M. Dewerchin, C. Molitor, D. Gheysen, M. Slaoui, and B. T. Rouse. 1995. Vaccination with recombinant vaccinia viruses expressing ICP27 induces protective immunity against herpes simplex virus through CD4⁺ Th1⁺ T cells. *J. Virol.* **69**:4711-4716.
5. Mercadel, C., S. M. Brown, M. Slaoui, and B. T. Rouse. 1993. Efficacy of herpes simplex virus types 1 and 2 mutant viruses to confer protection against zosteriform spread in mice. *Viral Immunol.* **6**:35-42.
6. Mester, J. C., J. C. Glorioso, and B. T. Rouse. 1991. Protection against zosteriform spread of herpes simplex virus by monoclonal antibodies. *J. Infect. Dis.* **163**:263-269.
7. Kapoor, A. K., A. A. Nash, P. Wildy, J. Phelan, C. S. McLean, and H. J. Field. 1982. Pathogenesis of herpes simplex virus in congenitally athymic mice: the relative roles of cell mediated and humoral immunity. *J. Gen. Virol.* **60**:225-233.
8. Killeen, N., S. Sawada, and D. R. Littman. 1993. Regulated expression of human CD4 rescues helper T-cell development in mice lacking expression of endogenous CD4. *EMBO J.* **12**:147-153.
9. Niemialowski, M., V. L. Godfrey, and B. T. Rouse. 1994. Quantitative studies on CD4⁺ and CD8⁺ cytotoxic T lymphocyte responses against herpes simplex virus type 1 in normal and $\beta 2M$ deficient mice. *Immunobiology* **190**:183-194.
10. Roizman, B., and A. E. Sears. 1990. Herpes simplex virus and their replication, p. 1795-1841. *In* B. N. Fields and D. M. Knipe (ed.), *Virology*. Raven Press, New York.
11. Schmid, D. S., and B. T. Rouse. 1992. The role of cytotoxic T lymphocytes in control of HSV. *Top. Microbiol. Immunol.* **179**:57-74.
12. Simmons, A., and A. A. Nash. 1984. Zosteriform spread of herpes simplex virus as a model of recrudescence and its use to investigate the role of immune cells in prevention of recurrent disease. *J. Virol.* **52**:816-821.
13. Simmons, A., and D. C. Tschärke. 1992. Anti-CD8 impairs clearance of herpes simplex virus from the nervous system: implications for the fate of virally infected neurons. *J. Exp. Med.* **175**:1337-1344.
14. Smith, P. M., R. M. Wolcott, R. Chervenak, and S. R. Jennings. 1994. Control of acute cutaneous herpes simplex virus infection: T-cell mediated viral clearance is dependent upon interferon- γ (IFN- γ). *Virology* **202**:76-88.
15. York, I. A., C. Roop, D. W. Andrews, S. R. Riddell, F. L. Graham, and D. C. Johnson. 1994. A cytosolic herpes simplex virus protein inhibits antigen presentation to CD8⁺ lymphocytes. *Cell* **77**:525-536.