Hydrolyzable Tannins (Chebulagic Acid and Punicalagin) Target Viral Glycoprotein-Glycosaminoglycan Interactions to Inhibit Herpes Simplex Virus Type 1 Entry and Cell-to-Cell Spread

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ABSTRACT

Herpes simplex virus type 1 (HSV-1) is a common human pathogen that causes life-long latent infection of sensory neurons. Non-nucleoside inhibitors that can limit HSV-1 recurrence are particularly useful in treating immunocompromised individuals or cases of emerging acyclovir-resistant strains of herpes virus. We report that chebulagic acid (CHLA) and punicalagin (PUG), two hydrolyzable tannins isolated from the dried fruits of Terminalia chebula Retz. (Combretaceae), inhibit HSV-1 entry at non-cytotoxic doses in A549 human lung cells. Experiments revealed that both tannins targeted and inactivated HSV-1 viral particles, and could prevent binding, penetration, cell-to-cell spread, as well as secondary infection. The antiviral effect from either of the tannins was not associated with induction of type I interferon-mediated responses, nor was pre-treatment of the host cell protective against HSV-1. Their inhibitory activities targeted HSV-1 glycoproteins since both natural compounds were able to block polykaryocyte formation mediated by expression of recombinant viral glycoproteins involved in attachment and membrane fusion. Our results indicated that CHLA and PUG blocked interactions between cell surface glycosaminoglycans and HSV-1 glycoproteins. Furthermore, the antiviral activities from the two tannins were significantly diminished in mutant cell lines unable to produce heparan sulfate and chondroitin sulfate, and could be rescued upon reconstitution of heparan sulfate biosynthesis. We suggest that the hydrolyzable tannins CHLA and PUG may be of value as competitors for glycosaminoglycans in the management of HSV-1 infections, and that they may help reduce the risk for development of viral drug resistance during therapy with nucleoside analogues.
INTRODUCTION

Herpes simplex virus type 1 (HSV-1) is an alpha-herpesvirus that typically causes mucocutaneous lesions in oral, perioral, and other mucosal sites in the body (58). The virus commonly uses the oropharyngeal mucosa as a port of entry, and following primary infection, establishes a lifelong latent state in the host’s trigeminal ganglia sensory neurons. Sporadic recurring infections occur when HSV-1 is reactivated by various stimuli, such as sunlight, immunosuppression, menstruation, fever, or stress (23). While primary or reactivated HSV-1 infections can be subclinical or manifested by mild and self-limited diseases, severe cases of this viral infection may lead to complications such as keratoconjunctivitis, meningitis, and encephalitis (3, 5). Importantly, corneal HSV-1 infection can lead to stromal keratitis, which remains one of the leading causes of blindness in developing countries (37). More aggressive diseases due to HSV-1 are common in immunocompromised individuals (3, 5, 23). To date, no treatment has been identified that eradicates or resolves latent infections by this ubiquitous pathogen.

HSV-1 viral entry into cells is initiated by interaction of viral envelope glycoproteins (gB and gC) with host cell surface proteoglycans (PGs) conjugated to glycosaminoglycans (GAGs) containing heparan sulfate (HS) or chondroitin sulfate (CS) moieties. These initial interactions are sufficient for viral adsorption but not viral entry (67). Subsequently, higher affinity interaction of gD with its receptors including herpesvirus entry mediator (HVEM, member of tumor necrosis factor [TNF] receptor family), nectin-1 and nectin-2 (two members of the immunoglobulin superfamily), and/or 3-O-sulfated HS, leads to fusion of the viral membrane with either the plasma or endosomal membranes of the cell through further interactions with gB, gH, and gL (29, 57, 67). Initial interaction of HSV-1 with GAGs ensures a highly efficient infection, but infection of cells deficient in HS or CS can still be achieved via the high affinity receptors, albeit at lower efficiency. Following transport of the viral capsid to the nucleus, where the HSV-1 genome is released, viral products are then expressed in a sequential and coordinated fashion, and are divided into three groups of virus-specific proteins designated as immediate early (α) (ICP0 and ICP4), early (β) (ICP8, UL42, and thymidine kinase), and late (γ) (gB, gC, gD, and gH) phase proteins (73). Although cellular innate immunity is activated upon virus infection, HSV-1 can produce one or more proteins that counteract the host antiviral response (46, 49, 50).

Most anti-herpes drugs target the viral DNA polymerase, and include nucleoside or pyrophosphate analogues. Acyclovir (ACV), a guanosine analogue, has been the most...
important clinical drug for the prophylactic or therapeutic treatment of HSV infections, and represents the gold standard for anti-HSV therapy (4, 62). However, extensive use of this drug has led to clinical problems with the emergence of ACV-resistant virus strains, particularly in immunocompromised patients, including those who have had transplantation surgery or have been infected by the human immunodeficiency virus (HIV) (9, 21, 24, 48, 70, 79). Subsequent management of ACV-resistant patients using a different class of DNA polymerase-targeting inhibitor, such as foscarinet, has also been hindered by drug resistance (21, 59). There is a need to identify alternative antiviral therapies with different modes of action to improve the treatment and control of HSV infections, especially in immunocompromised individuals.

*Terminalia chebula* Retz. (*T. chebula*), a member of the *Combretaceae* family, is a traditional medicinal plant that is native to India and Southeast Asian countries. The dried ripened fruit of *T. chebula* (Fructus Chebulae), often referred to as “myrobalans”, contains antioxidants (15) and is commonly used as a broad spectrum medicinal agent for the treatment of dysentery, asthma, cough, sore throat, bloody stools, and diseases of the heart and bladder (30). *T. chebula* is rich in tannins, which are polyphenolic secondary metabolites found in higher plants (27, 32, 36). Tannins are characterized by their relatively high molecular weight (500 to 20,000 daltons) and the unique ability to form insoluble complexes with proteins, carbohydrates, nucleic acids, or alkaloids (27, 55, 63). The hydrolyzable class of tannins possesses structures that generally consist of gallic or ellagic acid esters conjugated to a sugar moiety (28, 36). These polyphenols have high affinity for proteins and polysaccharides, and are thought to be the major bioactive compounds found in the leaves and the fruit of *T. chebula*.

Antiviral activities from hydrolyzable tannins are well documented and are generally thought to target viral adsorption to the host cell membrane (for HSV and HIV), as well as reverse-transcriptase activity of HIV (reviewed in (8) and (63)). We have previously identified several tannins of various plant sources that exhibit potent antiviral activities against HSV-1 and HSV-2. These include 1,3,4,6-tetra-O-galloyl-beta-D-glucose (77), casuarinin (10), *ent*-Epiafzelechin-(4α→R8)-epiafzelechin (17), excoecarianin (16), geraniin (77), hippomanin A (76), prodelphinidin B-2 3’-O-gallate (11), prodelphinidin B-2 3,3’-di-O-gallate (12), pterocarnin A (13), and putranjivain A (14). Studies from other laboratories have also reported a series of tannins and related compounds capable of inhibiting HSV infections (25, 56, 64, 71, 72). These earlier reports provide strong precedent.
for our studies and suggest that the tannins constitute an excellent focus for antiviral
discovery, particularly in the field of HSV therapeutics.

Identification of multiple drugs that can act on different phases of the viral life cycle can
be particularly useful in managing HSV-1 infection or reactivation in either
immunocompromised individuals, or cases of ACV resistance. To pursue this goal, we
extended our previous studies and concentrated our efforts on four chemically-defined
hydrolyzable tannins (Fig. 1), including chebulagic acid (CHLA), chebulinic acid (CHLI),
punicalagin (PUG), and punicalin (PUN), which are present in T. chebula (39, 40, 78).
Although an effect against HSV-1 has been previously reported for CHLA, the mechanism of
its activity was not elucidated (71). In this study we report that two of the tannins tested,
specifically CHLA and PUG, were found to be most effective against HSV-1. Detailed studies
into their inhibitory action revealed that both drugs specifically target HSV-1 particles, block
virus entry into the cell, inhibit cell-to-cell spread of the virus, and reduce secondary infection
from released virions. The antiviral mechanism is attributed to the binding of CHLA and
PUG to viral glycoproteins that interact with cell surface GAGs. Their ability to effectively
control viral entry and spread, underscore the potential of these two hydrolyzable tannins for
treating HSV-1 infection/recurrence.
MATERIALS AND METHODS

Chemicals and reagents
Dulbecco’s modified Eagle’s media (DMEM) and fetal calf serum (FCS) were supplied by Wisent Scientific (St-Bruno, QC, Canada). Gentamicin and fungizone were purchased from GIBCO-Invitrogen (Carlsbad, CA, USA). Acyclovir (ACV, Acycloguanosine) was obtained from Calbiochem (EMD Biosciences, Darmstadt, Germany). Foscarinet (FOS, Sodium phosphonoformate tribasic hexahydrate), dimethylsulfoxide (DMSO), and in vitro toxicology assay kit (XTT-based) were purchased from Sigma (St.Louis, MO, USA).

Test compounds
Fructus Chebulae and dried leaves from T. chebula were commercially obtained from Uchida Wakanyaku Co. (Tokyo, Japan) and an herbal market in Ping-Tung, Taiwan, respectively. Prior to extraction, both materials were anatomically authenticated by Chung-Ching Lin. CHLA, CHLI, and PUG were extracted from Fructus Chebulae, and PUN was derived from the leaves of T. chebula. The tannins were isolated and purified as described previously (39, 40). Before use, the structure of each compound was further confirmed by HPLC-UV/ESI-MS analyses, and their purities were checked using HPLC with photodiode array detection (HPLC-PDA) as previously reported (34, 35). CHLA, CHLI, PUG, PUN, and ACV were dissolved in DMSO. FOS was dissolved in sterile ddH₂O. All compounds were diluted with cell culture medium before use. The final concentration of DMSO in the drug solution was below or equal to 1% at the effective doses used.

Plasmids
The pCAGGS/MCS vector and its derivative plasmids expressing HSV-1 gB (pPEP98), gD (pPEP99), gH (pPEP100), and gL (pPEP101) (54) were generously provided by Drs. Patricia G. Spear and Richard Longnecker (Northwestern University, Chicago, IL, USA).

Cells and viruses
Vero (African green monkey kidney cells, ATCC CCL 81), HEL (Human embryonic lung fibroblast, ATCC CCL 137), and A549 (human lung carcinoma, ATCC CCL-185) cells were obtained from the American Type Culture Collection (ATCC; Rockville, MD, USA) and cultured in DMEM supplemented with 10% FCS, 50 µg/ml gentamicin, and 0.5 µg/ml fungizone at 37 °C in a 5% CO₂ incubator. Mouse L cells (provided by Dr. Bruce W.
Banfield, Queen’s University, Kingston, ON, Canada) and its mutant derivative cell lines gro2C (26) (obtained from Drs. Gary H. Cohen and Roselyn J. Eisenberg, University of Pennsylvania, Philadelphia, PA, USA) and sog9 (6) were cultured as above. Sog9-EXT1 cells were established as previously described (45) by transfecting sog9 cells with plasmid expressing the exostosin-1 (EXT1) gene and selecting in media containing 700 µg/ml G418. HSV-1 KOS strain (a gift from Dr. James R. Smiley, University of Alberta, Edmonton, AB, Canada), HSV-1 KOS strain with green fluorescent protein tag (HSV-1-GFP; provided by Dr. Karen L. Mossman, McMaster University, Hamilton, ON, Canada) (47), and vesicular stomatitis virus with green fluorescent protein tag (VSV-GFP; Indiana serotype, a gift from Dr. Brian D. Lichty, McMaster University, Hamilton, ON, Canada) (69) were propagated in Vero cells. HSV-1-GFP and VSV-GFP exhibit similar infectivity as their non-tagged wild-type counterparts. Virus titers were determined by standard plaque assay. Overlay media containing 0.1 % Gamunex (purified clinical human IgGs, provided by Dr. Andrew C. Issekutz, Dalhousie University, Halifax, NS, Canada) or 2 % methylcellulose were used for determination of viral titer for HSV-1 and VSV, respectively. The basal medium for the antiviral assays consisted of DMEM plus 2 % FCS with antibiotics.

Cytotoxicity assay

The cytotoxic effects of CHLA, CHLI, PUG, and PUN on the different cell types used in this study were measured by the calorimetric XTT (2,3-bis[2-methoxy-4-nitro-5-sulfophenyl]-5-phenylamino)-carbonyl]-2H-tetrazolium hydroxide) assay as described previously with some modifications (16). Briefly, cells were seeded in 96-well plates (1 × 10^4 cells per well) and incubated overnight to form a monolayer. Increasing concentrations of the test compounds were then applied to the culture wells in triplicate. After incubation at 37 °C for 72 h, the medium on the plate was discarded and the cells were washed twice with phosphate buffered saline (PBS). A volume of 100 µl assay solution from the in vitro XTT-based toxicology assay kit was added to each well. The plates were incubated for another 3 h to allow XTT formazan production. The absorbance was determined with an ELx800 Microplate reader (Bio-Tek Instrument, Inc.; Winooski, VT, USA) at a test wavelength of 492 nm and a reference wavelength of 690 nm. Data were calculated as percentage of surviving cells using the following formula: cell viability (%) = At / As × 100 %, where ‘At’ and ‘As’ refer to the absorbance of the test compounds and the solvent control (DMSO), respectively. The concentration of 50 % cellular cytotoxicity (CC50) of the test compounds was determined as the drug concentration that yielded 50 % cell death as previously described (19).
Antiviral plaque assay and drug dose-response analysis

A549 cells seeded in 12-well plates (5 × 10^5 cells per well) were treated with serial dilutions of the test compounds for 15 min at 37 °C, and then challenged with HSV-1 (50 PFU/well) for 1 h. The inoculums and drugs were subsequently removed from the wells, and the cells were washed with PBS twice and overlaid again with different dilutions of the test compounds. After further incubation for 48 h, the supernatant was removed, and the wells were fixed with methanol and stained with Giemsa stain solution (Sigma). Viral inhibition (%) was calculated as: \[1 - \text{(No. of plaques)}_{\text{exp}} / \text{(No. of plaques)}_{\text{control}}\] × 100 %, where (No. of plaques)_{exp} indicates the plaque counts from virus infection with test compound treatment and (No. of plaques)_{control} indicates the number of plaques derived from virus-infected cells with control treatment (HSV-1 with DMSO only) (10). The 50 % effective concentration (EC_{50}) for antiviral activity was defined as the concentration of antiviral compound that produced 50 % inhibition of the virus-induced plaque formation (19).

For dose-response determination, A549 cells seeded in 96-well plates were infected with HSV-1-GFP (MOI=1) in the presence or absence of the test compounds at various concentrations (0, 10, 20, 40, and 60 µM) for 24 h. The plates were then scanned by the Typhoon 9410 variable mode imager (Amersham Biosciences; Baie d’Urfe, QC, Canada) and the data was analyzed by ImageQuant TL software (Amersham Biosciences). Viral infection (%) was calculated as: \[(\text{Fluorescence})_{\text{exp}} - (\text{Fluorescence})_{\text{cell control}} / (\text{Fluorescence})_{\text{virus control}} - (\text{Fluorescence})_{\text{cell control}}\] × 100 %, where (Fluorescence)_{exp} indicates the GFP expression value from the virus-infected wells with drug treatment, (Fluorescence)_{cell control} signifies the GFP expression value of the cell control (DMSO only), and (Fluorescence)_{virus control} indicates the GFP expression value derived from virus-infected cells with control treatment (HSV-1-GFP with DMSO only). Values were obtained from three independent experiments with each sample assay performed in triplicate. A standard curve was also generated to ensure linear correlation between virus infection and GFP expression at the MOI assessed.

Assays for effect of tannin treatment at different times

The effect of drug addition over time was assessed according to a previously reported method with some modifications (41). To assess the effect of pre-treating cells with tannins, A549 cell monolayers seeded in 12-well plates were treated with CHLA (60 µM) and PUG (40 µM) for 24 h (long term) or 1 h (short term), and then washed with PBS before challenge with HSV-1.
(50 PFU/well) in DMEM containing 2 % FCS. To study the effect of adding tannins and virus concurrently, A549 cells were treated simultaneously with HSV-1 (50 PFU/well) and CHLA (60 µM) or PUG (40 µM). After incubation for 1 h at 37 °C, the virus-drug mixture was removed and cells were washed prior to overlay with media. To evaluate whether the tannins had any effects post viral entry, A549 cells were challenged with HSV-1 (50 PFU/well) for 1 h, and after removing the virus inoculum, infected cells were washed and subsequently overlaid with media containing CHLA (60 µM) or PUG (40 µM). For the continuous drug treatment, cells were pre-treated for 1 h with the tannins, challenged with HSV-1 in the presence of the drugs, and overlaid with media containing the test compounds after viral entry. For all the above experiments, viral plaques were stained and counted following a total incubation of 48 h post-infection (p.i.). DMSO (0.1 %) treatment was included as control in each condition.

Vesicular stomatitis virus (VSV) plaque reduction assay for host innate immunity
To evaluate whether CHLA and PUG induced host innate immune response, a VSV plaque reduction assay was performed. Briefly, A549 cells were seeded in 12-well plates (5 × 10^5 cells per well) and then pre-treated with CHLA (60 µM), PUG (40 µM), IFN-α from human leukocytes (1000 U/ml, Sigma), or with media and DMSO (0.1 %) controls for 24 h. Cell monolayers were washed with PBS twice and subsequently infected with VSV-GFP at MOI=0.01 for 1 h before applying the overlay media containing 2 % FCS and 2 % methylcellulose. The plates were scanned by Typhoon 9410 variable mode imager to visualize fluorescent plaques at 48 h p.i.

Viral inactivation assay
A viral inactivation assay was performed as previously described with some modifications (41). HSV-1 (1 × 10^4 PFU/ml) was mixed with CHLA (60 µM) or PUG (40 µM), and incubated at 37 °C for 1 h. The test compound-virus mixture was then diluted 50-fold (final virus concentration 50 PFU/well) with DMEM containing 2 % FCS to yield a sub-therapeutic concentration of the drug, and the virus inoculums were subsequently added to monolayers of A549 cells seeded in 12-well plates. As a comparison, HSV-1 was mixed with test compounds, diluted immediately to 50-fold (no incubation period) and added onto A549 cells for infection. The 50-fold dilution served to titrate the drugs below their effective doses and prevent meaningful interactions with the host cell surface. Following adsorption for 60 min at 37 °C, the diluted inoculums were discarded and cells were washed with PBS twice. An
overlay medium (DMEM containing 2 % FCS) was applied to each well and the plates were further incubated at 37 °C for 48 h before being subjected to plaque assay as described above. Viral plaques were counted and plaque numbers obtained from infections in the presence of drug compounds were compared to the 0.1 % DMSO control.

Viral attachment assay by ELISA and flow cytometry
Viral attachment to cell surface was assessed at 4 °C, which allows for HSV-1 binding but excludes entry (41), using cellular enzyme-linked immunosorbent assay (ELISA) as well as flow cytometry. The ELISA binding assay was performed as previously described with some modifications (2, 31). Briefly, 96-well plates were seeded with A549 cells (2 × 10^4 cells per well) and allowed to grow overnight. The cell monolayers were pre-chilled at 4 °C for 1 h and subsequently challenged with HSV-1 (MOI=5) inoculum in the presence of CHLA, PUG, or the DMSO (1 %) and heparin controls at various concentrations for 3 h at 4 °C. Cells inoculated with DMSO (1 %) alone served as mock control. Following infection, the wells were washed twice with ice-cold PBS to remove unbound virus, fixed with pre-chilled 4 % paraformaldehyde (PFA) in PBS for 1 h on ice, and then blocked with 5 % bovine serum albumin (BSA) at 4 °C. To detect bound virus by ELISA, samples were incubated at 37 °C for 1 h with a polyclonal rabbit anti-HSV-1 antibody (Dako Canada, Inc.; Mississauga, ON, Canada) at 1:7500 dilution in a PBS solution with 0.1 % Tween 20 (PBST) and containing 5 % BSA. The wells were subsequently washed two times with PBST containing 5 % BSA and two times with PBST only, with each wash performed at 5 min interval on a plate shaker to minimize background effects. The secondary antibody consisted of a goat anti-rabbit IgG conjugated with horseradish peroxidase (HRP; Invitrogen), and was added to the samples at 1:100000 dilution in PBST containing 5 % BSA. After incubation at 37 °C for 1 h, the wells were washed as previously described, and developed with the TMB (3,3',5,5'-tetramethylbenzidine) 2-component microwell peroxidase substrate kit (KPL; Gaithersburg, MD, USA) at room temperature for 20 min before stopping the reaction with 1 M phosphoric acid (H_3PO_4). The absorbance was immediately determined at 450 nm with an ELx800 Microplate reader. Values are expressed as fold change of absorbance relative to the mock infection control.

For the viral binding assay using flow cytometry analysis, A549 cells (1 × 10^6 cells per well) were first dissociated using a non-enzymatic cell dissociation buffer (Sigma). Cells were infected with HSV-1 (MOI=1) in the presence or absence of CHLA (60 µM) or PUG (40 µM) for 3 h at 4 °C. DMSO (0.1 %) was used as a negative control and heparin (100
µg/ml) was included as a positive control. Cells were subsequently washed twice with ice-cold FACS buffer (1×PBS, 2 % FCS, and 0.1 % sodium azide), blocked with 5 % FCS for 30 min on ice, and then stained with a fluorescein isothiocyanate (FITC)-conjugated polyclonal rabbit anti-HSV-1 antibody (1:500; Dako Canada, Inc). Stained samples were washed twice with FACS buffer and then fixed with 1 % PFA before being subjected to standard flow cytometry analysis. Normal rabbit serum was included in the experiments as an isotype control (1:250; Abcam). Data acquisition and flow cytometry analysis were performed on a Cyan flow cytometer (Dako Canada, Inc).

**Viral penetration assay**

The viral penetration assay was performed as previously described (41) with minor modifications. A549 cell monolayers grown in 12-well plates were pre-chilled at 4 °C for 1 h and subsequently incubated with HSV-1 (100 PFU/well) for 3 h at 4 °C to allow for viral adsorption. The infected cell monolayers were then incubated in the presence of CHLA (60 µM), PUG (40 µM), heparin (100 µg/ml), or DMSO (0.1 %) for an additional 20 min at 37 °C to facilitate HSV-1 penetration. At the end of the incubation, extracellular virus was inactivated by citrate buffer (pH 3.0) (18) for 1 min, and then cells were washed with PBS and overlaid with DMEM containing 2 % FCS. Following 48 h of incubation at 37 °C, viral plaques were stained and counted.

**Effect of tannin addition on viral RNA expression at different times post-entry**

The effects of tannin addition on HSV-1 RNA expression within the cell was performed by reverse-transcriptase (RT)-PCR analysis. A549 cells were infected with HSV-1 (MOI=1) for 1 h, and then treated with low pH citrate buffer (pH 3.0) to inactivate extracellular viral particles. Cells were then overlaid with media containing CHLA (60 µM), PUG (40 µM), or DMSO control (0.1 %). At 4, 8, and 12 h p.i., total cellular RNA was isolated using TRIzol Reagent (Invitrogen), treated with DNase I (Qiagen Inc.; Mississauga, ON, Canada) to remove genomic DNA, and purified by phenol/chloroform according to the protocols from the manufacturers. Aliquots of 1 µg RNA were used to generate cDNA with a High Capacity cDNA Reverse Transcription Kit (ABI; Foster City, CA, USA). The cDNA (10 %) was then subjected to standard PCR amplification using the following primers against HSV-1 immediate-early (ICP27), early (TK), and late (gD) genes, and also against the cellular glyceraldehyde-3-phosphate dehydrogenase (GAPDH) gene: ICP27 forward primer 5'-GCCGCGACGACCTGGAATCG-3', ICP27 reverse primer
5’-TGTGGGGCGCTGGTTGAGGATC-3’ (216 bp); TK forward primer
5’-AGGTATCGCGCGGCCGGGTAG-3’, TK reverse primer
5’-ATGGCTTCGTACCCCTGCCA-3’ (533 bp); gD forward primer
5’-ATGGGAGGCAACTGTGCTATCC-3’, gD reverse primer
5’-CTCGGTGCTCCAGGATAAAC-3’ (250 bp); GAPDH forward primer
5’-GCCTCCTGCACCACCAACTG-3’, GAPDH reverse primer
5’-ACGCCTGCTTCACCACCTTC-3’ (349 bp). For comparison, A549 cells were also infected with HSV-1 (MOI=1) in the presence of CHLA (60 µM), PUG (40 µM), or DMSO (0.1 %) at 0 time (co-addition). After incubation for 1 h at 37 °C, the inoculums and tannins were removed, and the cells were washed with PBS before being overlaid with DMEM containing 2 % FCS. Again at 4, 8, and 12 h p.i., total cellular RNA was harvested and subjected to RT-PCR analysis as described above.

Effect of tannins on HSV-1 secondary infection and cell-to-cell spread

To assess the effect of the tannins on secondary viral infections, confluent A549 cells were seeded in 12-well plates and then infected with HSV-1-GFP (200 PFU/well) for 1 h. After virus adsorption and penetration, extracellular virus was inactivated by citrate buffer (pH 3.0) for 1 min followed by PBS washes. The cells were then treated with CHLA (60 µM), PUG (40 µM), heparin (100 µg/ml), or DMSO (0.1 %) following one round of HSV-1 replication (12 h). Any increase in plaque formation due to released virions from secondary infections could be monitored over the subsequent incubation period. At 24 and 48 h post the initial infection, the plates were scanned using a Typhoon 9410 variable mode imager and fluorescent viral plaques were counted using the ImageQuant TL software to determine whether any new plaques were formed due to virus progeny. Values were obtained from three independent experiments with each sample being performed in duplicate and plotted on a bar graph.

For the analysis of virus cell-to-cell spread, experiments were performed as described above, except that the drugs were added to HSV-1-GFP-infected-cells at 24 h p.i. The overlay media also contained neutralizing Gamunex antibodies (0.1 %), which were used to prevent secondary infection from released HSV-1 virions. The antibodies do not affect lateral spread of virus between cells via intercellular junctions. This allowed the monitoring of cell-to-cell spread of virus in the presence and absence of tannins. Drugs were added at 24 h p.i. and a visual comparison of viral plaque size was performed at the end-point of the experiment (48 h p.i.). Photomicrographs were taken at 100× magnification (Leica Microsystems; Wetzlar,
Germany) between 24 and 48 h p.i.

**Virus-free cell fusion assay**

To examine whether the compounds interacted with HSV-1 glycoproteins to inhibit glycoprotein-mediated fusion events, a virus-free cell fusion assay was performed (54). A549 cells were seeded in 6-well dishes and transfected with plasmid DNA expressing the individual HSV-1 glycoproteins (gB, gD, gH, and gL) or with the control vector using Arrest-In (Open Biosystems; Huntsville, AL, USA). The total amount of plasmid DNA per well was 4 µg. After 6 h of incubation at 37 °C, the transfection mixture was discarded and the cells were washed with PBS before treatment with CHLA (60 µM), PUG (40 µM), heparin (100 µg/ml), or DMSO (0.1 %). After incubation for another 24 h, cells were fixed with methanol and then stained with Hoescht dye (Sigma) or Giemsa stain solution. Photomicrographs were then taken at 200× magnification (Leica Microsystems). Polykaryocytes with > 10 nuclei from the dishes were also counted.

**Effect of tannins using plaque assays in GAGs-deficient cell lines**

For antiviral analysis in GAGs mutant cell lines, confluent mouse L, gro2C, sog9, and sog9-EXT1 cells in 12-well dishes were infected with HSV-1-GFP, at 200 PFU/well (MOI=0.0004) reflecting viral titers obtained in each cell line, in the presence of CHLA (60 µM), PUG (40 µM), or DMSO (0.1 %). Following incubation for 1 h at 37 °C, the virus-drug mixture was removed and the cells were washed with PBS before overlaying with DMEM containing 2 % FCS. After further incubation for 48 h, fluorescent plaques were scanned as described above and counted using the ImageQuant TL software. Data was expressed as % viral infection = [(No. of plaques)exp / (No. of plaques)control] × 100 %.
RESULTS

Inhibition of HSV-1 infection by the hydrolyzable tannins

Chebulagic acid (CHLA), chebulinic acid (CHLI), punicalagin (PUG), and punicalin (PUN) (Fig. 1) have been reported to exhibit antiviral activities. We investigated whether these hydrolyzable tannins could inhibit HSV-1 infection. In order to ensure that the tannin concentrations were not cytotoxic, a toxicity analysis was carried out in A549 cells by using the XTT cell viability assay. Our results indicated that these four tannins did not have apparent cytotoxic effects below 100 µM in A549 cells while a dose-dependent cytotoxic effect was seen when concentration > 100 µM was used (data not shown). The 50 % cellular toxicity indices (CC$_{50}$) of CHLA, CHLI, PUG, and PUN were 316.87 ± 9.01, 330.83 ± 9.07, 318.84 ± 4.98, and 310.85 ± 1.99 µM, respectively (summarized in Table 1). For comparison, toxicity in primary human fibroblasts (HEL) cells was also assessed, and similar results were observed (data not shown).

We then evaluated the antiviral effects of these four natural compounds against HSV-1 infection using a plaque assay. Acyclovir (ACV) and foscarnet (FOS) were used as positive controls and DMSO was included as a negative control. FOS is the treatment of choice in the clinical setting if resistance develops against ACV. All four tannins could inhibit viral plaque formation, following inoculation of 50 PFU, in a dose-dependent manner, and their 50 % effective concentration (EC$_{50}$) values were 17.02 ± 2.82 (CHLA), 20.85 ± 2.40 (CHLI), 10.25 ± 1.13 (PUG), and 21.33 ± 1.77 µM (PUN) (Table 1). The selectivity index (SI), which measures the preferential antiviral activity of a drug in relation to its cytotoxicity, was calculated according to their CC$_{50}$ and EC$_{50}$. The SI indices of CHLA, CHLI, PUG, and PUN were 18.62, 15.87, 31.11, and 14.57, respectively (Table 1). Given their higher SI values, CHLA and PUG were chosen for subsequent analyses.

To obtain a more accurate dose-response curve for these two hydrolyzable tannins, A549 cells were infected with HSV-1-GFP (MOI=1) in the presence of the tannins, and fluorescent signals were quantified. The HSV-1-GFP was susceptible to the antiviral effects of the tannins. Both CHLA and PUG displayed anti-HSV-1 activity in a dose-dependent manner (Fig. 2), and the concentrations of CHLA at 60 µM and PUG at 40 µM, which provided near complete protection against the virus infection at a MOI=1, were chosen for all subsequent experiments.

Antiviral activities of CHLA and PUG depend upon the presence of HSV-1, and
inhibition is not due to activation of host cell innate immunity
To better understand the antiviral mechanism and the stage of HSV-1 infection affected by these two T. chebula tannins, we added the compounds at different times of the virus life-cycle (pre-entry, entry, and post-entry). In order to study pre-entry, more specifically the effect of the compounds on the cell itself prior to virus addition, A549 cells were pre-treated with CHLA and PUG for long term (24 h) or short term (1 h) periods and then washed prior to HSV-1 infection. For effects on the viral entry stage, virus and the drugs were simultaneously applied to the cells. To investigate events following virus entry, A549 cells were first infected with HSV-1 for 1 h and then treated with the tannins. For comparison, the tannins were also maintained throughout the entire experimental period.

Pre-treating A549 cells with CHLA and PUG (both long term and short term) did not protect against HSV-1 infection. Both tannins were effective in preventing plaque formation when added during virus adsorption, immediately after viral entry, and throughout multiple cycles of virus replication (Fig. 3). The data indicate that HSV-1 infection is severely impaired only if the drugs are present at the time of infection or during viral spread, and that it is unlikely that the antiviral activity is due to direct effects on the cells (such as masking cellular receptors or entry factors for HSV-1). To confirm that neither of the tannins activated host cell innate immunity and induced production of antiviral cytokines such as interferons (IFNs), a VSV plaque reduction assay was performed. VSV replication is extremely sensitive to cellular IFN production. IFN-α, a potent inhibitor of VSV, was included as a positive control. In line with the above observation, neither CHLA nor PUG pre-treatments protected A549 cells from VSV infection, whereas IFN-α treatment produced an intact cell monolayer (Fig. 4). Moreover, neither of the tannins induced IFN-stimulated genes in the A549 cells (data not shown). The above results suggest that the anti-HSV-1 activities produced by CHLA and PUG: (1) are unlikely to be mediated by effects through binding to the cellular receptors for HSV-1 or in triggering antiviral innate immunity, and (2) absolutely requires the presence of HSV-1.

CHLA and PUG block HSV-1 entry by inactivating virus particles and preventing virus attachment and penetration into A549 cells
In order to evaluate the antiviral mechanism of CHLA and PUG, we investigated their effects on the virus particles themselves. Tannin compounds were pre-incubated with virus particles and then diluted to a sub-therapeutic concentration prior to infection. Both CHLA and PUG could interact with virus particles, irreversibly, to prevent infection (Fig. 5A). This suggests
that CHLA and PUG can bind to virus particles and neutralize virus infectivity.

We next assessed the ability of CHLA and PUG to affect viral attachment and penetration. The attachment assay was carried out at 4 °C, which allows for virus binding but prevents entry (41). Using ELISA to detect bound virus on the adherent cells, both tannin compounds were observed to inhibit HSV-1 attachment to the A549 cell surface in a dose-dependent manner (Fig. 5B). An alternative binding assay was also performed using virus-specific antibodies to detect bound HSV-1 particles by flow cytometry. Again, CHLA and PUG could prevent HSV-1 binding to the surface of the target A549 cells (Fig. 5C). Heparin, a competitive HSV binding inhibitor, was included as a positive control in both experiments. These results suggest that CHLA and PUG might interact with viral glycoprotein(s) and/or cellular receptor(s) during the virus attachment phase. To further assess the effects of CHLA and PUG on virus penetration step, HSV-1 particles were allowed to first bind to A549 cells at 4 °C, and were subsequently allowed to fuse with and penetrate the host cell membrane by shifting the temperature to 37 °C in the presence or absence of the tannins. As shown in Fig. 5D, CHLA retained most of its antiviral activity even during the viral penetration phase and PUG could completely abrogate virus penetration into the A549 cells, resulting in a protected monolayer. In contrast, heparin, which is effective at blocking HSV-1 adsorption, did not prevent subsequent virus penetration (42). The data indicate that CHLA and PUG impair viral receptor attachment and penetration functions during the HSV-1 infection.

CHLA and PUG do not affect HSV-1 transcription following entry but limit secondary viral infection and cell-to-cell transmission

The observation in Fig. 3, that CHLA and PUG inhibited HSV-1 plaque formation when treatment was initiated immediately after the virus had entered the cell, suggested that the two tannins may block HSV-1 replication cycle or inhibit HSV-1 secondary infection and/or cell-to-cell spread in the ensuing incubation period. To specifically address these possibilities, we first evaluated the effects of CHLA and PUG on HSV-1 mRNA expression following virus entry. A549 cells were infected with HSV-1 for 1 h, then extracellular virus was inactivated by citrate buffer treatment and washed away, and CHLA, PUG, or DMSO was subsequently added to the cells. For comparison, CHLA and PUG were also added simultaneously with HSV-1. Total cellular RNA was isolated from all samples at various time points following viral infection. Our results clearly indicated that CHLA and PUG did not affect HSV-1 mRNA expression following virus penetration, since levels of immediate-early
(ICP27), early (TK), and late (gD) viral gene transcripts were unaffected by the compounds (Fig. 6A). On the other hand, both tannins suppressed HSV-1 mRNA synthesis when added together with the virus at the same time (Fig. 6B). These findings suggest that neither CHLA nor PUG inhibit HSV-1 transcription following penetration of the host cell.

We next examined whether CHLA and PUG inhibited HSV-1 secondary infection and/or cell-to-cell spread. A fluorescent plaque assay was performed using HSV-1-GFP. After viral inoculation of A549 cells, CHLA and PUG were added to the overlay media at 12 or 24 h p.i. in the presence or absence of HSV-1 neutralizing antibodies, and fluorescent viral plaques were quantified or photographed over the subsequent course of infection (total of 48 h post initial challenge). In this assay, neutralizing antibodies coat viral particles released from infected cells and prevent secondary infection of surrounding uninfected cells. Thus, the only route of cell-to-cell transmission in the presence of neutralizing antibody is via intercellular junctions between infected and uninfected cells. Furthermore, prior to drug addition (12 and 24 h p.i.), HSV-1 was permitted to complete at least one round of replication cycle, allowing drug effects on post-entry infection to be monitored in the ensuing incubation period. As expected, in the absence of neutralizing antibodies, there was an increase in viral plaques in the DMSO control group due to secondary infections (Fig. 7A). However, addition of CHLA, PUG, or heparin substantially reduced the number of viral plaques formed and limited its increase in comparison to DMSO (Fig. 7A). Similarly, with respect to viral spread, treatment with CHLA and PUG prevented viral plaque growth. Incubation with the tannins yielded plaques with considerably reduced size compared to the DMSO control after the 48 h of infection (Fig. 7B). Interestingly, heparin, which has limited inhibitory activity on HSV-1 post-attachment, also exhibited some protective effect against viral spread albeit at a lower efficiency compared to the tannins. Taken together, the data indicated that once HSV-1 entered the cells and completed at least one cycle of infection (12-24 h), any subsequent de novo infections and viral spread via intercellular contacts were restricted upon addition of CHLA and PUG. Tannin-mediated inhibition of viral attachment and fusion, as observed earlier (Fig. 5B-D), confirmed these results, and may be responsible for their effects in neutralizing secondary infections and restricting cell-to-cell spread, respectively.

CHLA and PUG target HSV-1 glycoproteins that mediate glycosaminoglycan-specific interactions

HSV-1 viral glycoproteins are known to mediate HSV-1 binding, internalization, and cell-to-cell spread. From the preceding data, it appears that the hydrolyzable tannins CHLA

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and PUG target viral glycoprotein(s) which would explain the need for virus to be present during inhibition and their effect on virus entry and spread. In an attempt to elucidate the underlying mechanism, we checked whether the two tannins interacted with HSV-1 glycoproteins in order to block entry-associated events. Using a virus-free system, we overexpressed HSV-1 glycoproteins that have been shown to mediate cell fusion (occurs during entry and cell-to-cell spread) by transfecting the individual gB, gD, gH, and gL genes into A549 cells followed by treatment with the tannins. Expression of all four genes induced cell fusion resulting in polykaryocyte formation (> 10 nuclei) which is absent following transfection with the empty vector control. The two tannins and heparin each blocked polykaryocyte formation, suggesting that CHLA and PUG interact with HSV-1 glycoproteins to prevent virus attachment, entry, and cell-to-cell spread (Fig. 8).

Several HSV-1 glycoproteins are known to interact with cell surface GAGs. To further explore the virus-host interactions that are being targeted by the tannins, we used a series of cell lines known to possess defects in surface HS and CS synthesis. The relative infectivity of HSV-1 (KOS) is approximately 10% for HS-deficient gro2C cells and 0.5% for HS/CS-deficient sog9 cells compared to parental mouse L cells (6). Stable expression of the EXT1 gene in sog9 cells (sog9-EXT1) restores HS biosynthesis and susceptibility to HSV-1 infection (43). To evaluate the effects of the drugs in the presence or absence of GAG expression, each cell line was infected with different dilutions of HSV-1 sufficient to achieve 200 PFU/well (MOI=0.0004) in the presence of the tannins. CHLA and PUG effectively protected the parental mouse L cells and sog9-EXT1 cells from infection, but antiviral effects were diminished in HS-deficient gro2C cells, and almost completely abolished in HS/CS-deficient sog9 cells (Fig. 9). Similar results were obtained in experiments using different MOIs (data not shown). These observations strongly suggest that CHLA and PUG target interactions between HSV-1 glycoproteins and GAGs. CHLA inhibition also appeared to be more sensitive to cell surface deficiency in GAGs compared to that of PUG.

Taken together, the data indicate that CHLA and PUG function as GAG competitors to inhibit the initial events of HSV-1 infection (adsorption, penetration) and the cell-to-cell spread of virus. The interaction of HSV-1 glycoproteins with cellular GAGs plays a critical role in viral infections, and the hydrolyzable tannins could offer a primary means of defense against HSV-1 infections.
DISCUSSION

There is currently no cure that completely resolves latent infections caused by alpha-herpesviruses. Therefore, the development of small molecules capable of inhibiting infection by reactivated virus represents an attractive therapeutic strategy, particularly in immunocompromised individuals who are often at risk of generating ACV-resistant HSV-1 strains. In a search for such molecules, we report that CHLA and PUG, two hydrolyzable tannins isolated from the fruits of *T. chebula*, effectively inhibited HSV-1 infection in A549 cells, without significantly reducing cell viability. In addition, our results suggest that CHLA and PUG specifically targeted HSV-1 particles by binding to viral glycoproteins that interact with cellular GAGs, rendering the virus incapable of adsorbing, penetrating, and spreading throughout the cell monolayer. These features underscore the potential of tannins as HSV-1 entry inhibitors.

Our data show that entry events, including primary/secondary infection, viral attachment/penetration, and cell-to-cell spread are inhibited only when the tannins and HSV-1 glycoproteins are in contact with each other. Pre-treatment of host cells with the tannins, followed by washes to remove unadsorbed compounds, had no effects upon HSV-1 replication. This indicated that masking cell surface receptors or entry factors for HSV-1 by the tannins is unlikely. Viral binding assays using ELISA and flow cytometry analyses revealed that the tannins blocked viral attachment to the host cell. While CHLA and PUG could inactivate the HSV-1 particles, we do not believe that a direct lysis effect of the viral membrane is responsible for their effects, since HSV-1 infection of GAG-deficient mutant cell lines was still observed, even in the presence of these compounds (Fig. 9). Given their large molecular weight (CHLA = 954 MW and PUG = 1084 MW) and high affinity for proteins and sugars, the two hydrolyzable tannins are thought to bind to HSV-1 glycoproteins on the infectious virions making them inert, impairing glycoprotein function, and preventing successful attachment and entry of the host cell. These tannins could also bind to viral glycoproteins on the infected-cell surface, rendering them unavailable to mediate the cell-to-cell spread of virus.

HSV-1 entry into epithelial cells, which express the cellular receptors (HS, HVEM, nectin-1, and nectin-2) for HSV (67), requires an ordered and concerted effort from the viral glycoproteins. Specifically, the glycoproteins gB, gC, gD, gH, and gL interact with host cell receptors and are involved in penetration of the plasma membrane through a membrane fusion process (29, 57, 67). While viral entry and spread require a particular combination of
viral surface proteins, several HSV-1 glycoproteins are repeatedly involved in both processes. Importantly, gB and gD function in viral binding and fusion, and are also engaged during cell-to-cell transmission in cultured epithelial cells (22, 29, 33, 38, 57, 60, 61, 67). The associations between viral glycoproteins that mediate HSV-1 attachment and entry represent a complex scenario when considering CHLA and PUG and their mechanism of action. The candidate targets of the tannins likely involve viral glycoproteins that interact with host cell surface GAGs and participate in adsorption, membrane fusion, and cell-to-cell spread. The observation that both tannins blocked virus attachment to cells, as did heparin, suggests that interaction of gC and gB with heparan sulfate proteoglycans (HSPGs) is targeted. However, the drugs also prevented virus internalization into cells in the post-binding phase. At this point, the interaction with HSPGs should be irrelevant, since virions now interact with a gD receptor and become resistant to removal by heparin (42) (Fig. 5D). One explanation is that the tannins bind to gB and block its interaction with HSPG while also interfering with its subsequent role in membrane fusion during virus entry (in which gC is not involved). Alternatively, the tannins may be impeding the activity of additional glycoproteins (gD and/or gH/gL) or working by some other mechanism(s) to prevent successful entry into the A549 cells. Finally, there is the possibility that viral glycoproteins may still be accessible to the tannins, even when these viral proteins are bound to the host cell or are expressed in the intercellular junctions. This could explain why the considerably larger heparin (17,000-19,000 MW) can bind free gB, but is unable to interact efficiently with the shielded glycoprotein which is needed in order to inhibit viral penetration or cell-to-cell transmission. Additional binding experiments using glycoprotein-deficient HSV-1 mutants as well as soluble recombinant HSV-1 glycoproteins could help elucidate the targeting specificity of the tannins. We speculate that the two natural compounds can bind to all GAG-interacting glycoproteins including gB, gC, and/or gD, and neutralize their functions. The ability of CHLA and PUG to effectively block virus membrane penetration, as well as virus attachment, could explain their higher efficacy in restricting the spread of HSV-1 compared to the inhibitory effects of heparin.

In the case of HSV-1, the interactions between several of its glycoproteins and cell surface GAGs are critical for ensuring efficient viral entry as well as viral spread (6, 26, 44, 52, 53, 65, 66, 68). CS can confer susceptibility to HSV-1 infection in the absence of HS (7, 26), but the HS is still the preferred substrate for viral attachment. GAG deficiency renders cell surfaces relatively resistant to HSV-1 binding and residual infectivity relies on stable attachment receptors (6). Earlier studies have shown that the HS/CS-deficient sog9 cells are
insensitive to inhibitory effect of soluble HS on HSV-1 infection (6). We observed that while
the absence of HS in gro2C cells weakened the tannins’ inhibitory effects, the absence of HS
and CS on sog9 cells severely limited their antiviral activities (Fig. 9). Overexpression of the
EXT1 gene, which restores HS biosynthesis in EXT1 mutant sog9 cells, rescued the antiviral
effects from both CHLA and PUG to > 90%. The results suggest that the tannins interfere
with GAG interactions with viral glycoproteins which can involve attachment and may also
affect downstream penetration of the host cell. Additional experiments are required to clarify
whether the drugs are able to inhibit binding and/or fusion of HSV-1 on the GAG-deficient
cells.

All four hydrolyzable tannins investigated in this report are composed of a
glucopyranose core linked with galloyl derivatives, including hexahydroxydiphenoyl (HHDP; C-C coupling between galloyl moieties), gallagyl, and chebuloyl units (Fig. 1) (39, 78). Only
CHLA and PUG possess the HHDP unit, with an (R) configuration (linked to the glucose core
at the 3,6-positions) and (S) configuration (linked via the 2,3- or 4,6-positions of their glucose
residue), respectively (36, 78). The anti-herpes activities of hydrolyzable tannins are thought
to be dependent on the number of galloyl or HHDP groups, irrespective of the sugar core (71).
Structures containing HHDP unit have also been valuable pharmacophores for inhibiting HIV
enzymatic activities (20, 75). Indeed, anti-HIV activities from CHLA, CHLI, PUG, and PUN
have been reported to prevent binding of recombinant HIV gp120 to CD4 and to exert
inhibitory effects on HIV-1 RT and integrase (1, 51, 74). The ability of these natural agents to
inhibit both HSV-1 and HIV-1 underscores their potential value in the treatment of acquired
immunodeficiency syndrome patients who also exhibit HSV-1-related symptoms.

Use of these tannins could improve the prognosis of anti-HSV-1 therapy in
immunosuppressed individuals and help to reduce the risk of ACV-resistance by lowering the
ACV-dose required. Since Fructus Chebulae contains both CHLA and PUG, inclusion of
purified extracts from this plant in topical creams or microbicides would be a feasible method
for controlling recurrent HSV-1 infections. Future studies will determine whether these
natural products are effective against additional members of the herpesvirus family and other
enveloped viruses. Our preliminary studies have shown that both CHLA and PUG inhibit the
Other viruses known to use GAGs as entry factors, such as measles virus and human
respiratory syncytial virus, are also inhibited by these tannins, reinforcing our discovery that
these compounds act as GAG competitors that inhibit viral glycoprotein-cell receptor
interactions (Lin, L.-T. and Chen, T.-Y., unpublished data). Further studies with the tannins
derived from *T. Chebula* may provide new ways to inhibit recurrent HSV-1 infections and control the re-emergence of this virus in immunocompromised patients.

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AUTHOR CONTRIBUTIONS

Conceived and designed the experiments: LTL TYC CCL CDR. Performed the experiments: LTL TYC. Analyzed the data: LTL TYC CM CCL CDR. Contributed reagents/materials/technical support: LTL TYC CYC RSN CM T BG TCL GHW CCL CDR. Wrote the paper: LTL TYC CCL CDR.
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### TABLE 1

Cytotoxicity and anti-HSV-1 activity of CHLA, CHLI, PUG, and PUN in A549 cells.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>CC&lt;sub&gt;50&lt;/sub&gt;&lt;sup&gt;a&lt;/sup&gt; (µM)</th>
<th>Anti-HSV-1 effects</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>EC&lt;sub&gt;50&lt;/sub&gt;&lt;sup&gt;b&lt;/sup&gt; (µM)</td>
</tr>
<tr>
<td>CHLA</td>
<td>316.87 ± 9.01</td>
<td>17.02 ± 2.82</td>
</tr>
<tr>
<td>CHLI</td>
<td>330.83 ± 9.07</td>
<td>20.85 ± 2.40</td>
</tr>
<tr>
<td>PUG</td>
<td>318.84 ± 4.98</td>
<td>10.25 ± 1.13</td>
</tr>
<tr>
<td>PUN</td>
<td>310.85 ± 1.99</td>
<td>21.33 ± 1.77</td>
</tr>
<tr>
<td>ACV</td>
<td>&gt; 2000</td>
<td>14.45 ± 1.71</td>
</tr>
<tr>
<td>FOS</td>
<td>&gt; 2000</td>
<td>183.37 ± 25.18</td>
</tr>
</tbody>
</table>

<sup>a</sup> Cytotoxic effects were evaluated by XTT assay to determine the concentration of 50% cellular cytotoxicity (CC<sub>50</sub>) of test compounds.

<sup>b</sup> Antiviral effects were evaluated by plaque assay to determine the effective concentration that achieved 50% inhibition (EC<sub>50</sub>) against HSV-1 infection.

<sup>c</sup> Selectivity index (SI) = CC<sub>50</sub> / EC<sub>50</sub>.

Values shown are mean ± SEM (standard error of the mean) from three independent experiments with each treatment in triplicate.
FIG. 1. Chemical structures of chebulagic acid (CHLA), chebulinic acid (CHLI), punicalagin (PUG), and punicalin (PUN). Components of the tannins including galloyl, hexahydroxydiphenoyl (HHDP), gallagyl, and chebuloyl units are indicated.
FIG. 2. Dose-response for inhibition of HSV-1 infection in A549 cells by CHLA and PUG. A549 cells were seeded into 96-well plates and then infected with HSV-1-GFP (MOI=1) in the presence or absence of the tannins at various concentrations (0, 10, 20, 40, and 60 µM) for 24 h. DMSO (0.1 %) served as negative control. Viral infection was quantified by measuring GFP fluorescence using a Typhoon 9410 variable mode imager. Data shown are the mean ± SEM of three independent experiments with each tannin treatment being performed in triplicate.
FIG. 3. Effect of time of CHLA and PUG addition on plaque formation by HSV-1. A549 cells were seeded in 12-well plates and then treated with CHLA (60 µM) or PUG (40 µM) at various stages of HSV-1 infection (50 PFU/well). For pre-treatment, A549 cells were incubated with the test compounds for 24 h (long term) or 1 h (short term) and then washed once before infecting with HSV-1. For the co-addition experiment, the tannins and virus inoculum were added simultaneously to cells, incubated for 1 h, and then washed. In the post-infection experiment, the cells were infected with HSV-1 for 1 h, washed, and then subsequently treated with the tannins. In addition, cells were also incubated in the continuous presence of the test compounds from pre-treatment to post-infection stages. After an additional 48 h of incubation, viral plaques were stained and counted. DMSO (0.1 %) was included as control. Data shown are mean ± SEM of three independent experiments with each treatment performed in duplicate.
FIG. 4. CHLA and PUG do not induce IFN-mediated immune response against VSV infection. A549 cells were seeded in 12-well plates and then pre-treated with CHLA (60 µM), PUG (40 µM), IFN-α (1000 U/ml), or with media and DMSO (0.1 %) controls for 24 h. Cell monolayers were washed with PBS twice and then infected with VSV-GFP (MOI=0.01) for 1 h before applying overlay media containing 2 % FCS and 2 % methylcellulose. After an additional 48 h of incubation, the plates were scanned for visualization of fluorescent plaques. Data shown are representative images from one of two independent experiments.
FIG. 5. CHLA and PUG inhibit HSV-1 entry by inactivating viral particles and preventing virus binding and internalization into A549 cells. (A) Viral inactivation assay. HSV-1 (1 × 10^4 PFU/ml) was mixed with CHLA (60 µM) or PUG (40 µM) for 1 h at 37 °C, and then diluted 50-fold (final virus concentration 50 PFU/well) before infecting A549 cells. As a control, the same amount of virus was also mixed with the tannin, but diluted immediately and applied to the A549 cells. After a 48 h incubation period, viral plaques were stained and counted. DMSO (0.1 %) was included as a negative control. Data shown are mean ± SEM of three independent experiments with each treatment performed in duplicate. (B) Viral attachment analysis using ELISA. Adherent A549 cell monolayers in 96-well plates were pre-chilled at 4 °C for 1 h and then inoculated with HSV-1 (MOI=5) in the presence of CHLA, PUG, or the DMSO (1 %) and heparin controls at various concentrations for another 3 h at 4 °C. Wells were washed to remove unadsorbed virus, subsequently fixed with 4 % PFA, and then blocked with 5 % BSA. ELISA was performed using a primary polyclonal rabbit anti-HSV-1
antibody (1:7500) and a secondary goat anti-rabbit IgG conjugated with HRP (1:100000), followed by development with TMB substrate and reaction termination with 1 M H$_3$PO$_4$. The absorbance was immediately determined at 450 nm. Values are expressed as fold change of absorbance relative to the mock infection control (cells + 1 % DMSO) which is indicated by the dashed line. Data shown are mean ± SEM of three independent experiments with each treatment performed in triplicate. (C) Viral binding assay using flow cytometry analysis. Dissociated A549 cells were infected with HSV-1 (MOI=1) in the presence or absence of 60 µM CHLA or 40 µM PUG for 3 h at 4 °C. Cells were then washed, blocked, and stained with FITC-conjugated polyclonal rabbit anti-HSV-1 antibody (1:500). Stained samples were washed and fixed with 1 % PFA before being subjected to standard flow cytometry analysis. DMSO (0.1 %) was used as a negative control and heparin (100 µg/ml) was included as a positive control. Color indication for different treatments: grey = mock + DMSO, red = HSV-1 + DMSO, blue = HSV-1 + CHLA, green = HSV-1 + PUG, purple = HSV-1 + heparin. Data shown are representative of three independent experiments. (D) Viral penetration analysis by plaque assay. A549 cells were pre-chilled at 4 °C for 1 h before inoculation with HSV-1 (100 PFU/well) for 3 h at 4 °C. Then, cells were treated in the presence or absence of CHLA (60 µM), PUG (40 µM), or heparin (100 µg/ml), and further incubated for an additional 20 min with the temperature shifted to 37 °C to facilitate viral penetration. At the end of the incubation, extracellular virus was inactivated by citrate buffer (pH 3.0) for 1 min, then washed with PBS twice before overlaying with medium. After 48 h of incubation at 37 °C, viral plaques were stained and counted. DMSO (0.1 %) was included as negative control. Data shown are mean ± SEM of three independent experiments with each treatment performed in duplicate.
FIG. 6. CHLA and PUG do not affect HSV-1 transcription following entry into the host cell. (A) A549 cells were infected with HSV-1 (MOI=1) for 1 h, treated with low pH citrate buffer (pH 3.0) to inactivate non-internalized extracellular viral particles, and subsequently overlaid with media containing CHLA (60 μM), PUG (40 μM), or DMSO control (0.1 %). At 4, 8, and 12 h p.i., total cellular RNA was isolated, subjected to first-strand synthesis by reverse-transcription, and then amplified by standard PCR procedures with primers against HSV-1 immediate-early (ICP27), early (TK), and late (gD) gene products. GAPDH was included for control. (B) A549 cells were co-incubated with HSV-1 (MOI=1) and CHLA (60 μM), PUG (40 μM), or DMSO control (0.1 %) for 1 h. Cells were washed with PBS before applying overlay media without the tannins. Total cellular RNA was isolated for RT-PCR analysis as in (A). Representative data shown are from one of two independent experiments.
FIG. 7. CHLA and PUG can limit HSV-1 secondary infection and cell-to-cell spread of the virus. A549 cells were infected with HSV-1-GFP (200 PFU/well) for 1 h, then treated with citrate buffer (pH 3.0) to inactivate non-internalized extracellular viral particles. Cells were overlaid with (A) media or (B) media containing 0.1% neutralizing antibody. Following an incubation period of 12 h (A) or 24 h (B) p.i., infected cells were treated with CHLA (60 µM), PUG (40 µM), heparin (100 µg/ml), or DMSO (0.1%), before further incubating for a total of 48 h post the initial infection. Over the course of infection subsequent to the drug addition, the plates were (A) scanned and quantified for fluorescent viral plaques or (B) photographed using an inverted fluorescence microscope at 100× magnification. (A) Number of fluorescent plaques counted between 24 and 48 h p.i. with drug treatment initiated at 12 h post viral challenge. Data shown are mean ± SEM of three independent experiments with each treatment performed in duplicate. (B) Comparison of viral plaque size between 24 and 48 h p.i. with drug treatment initiated at 24 h post viral challenge; scale bars: 100 µm. Representative images are from one of two independent experiments.
FIG. 8

CHLA and PUG can prevent HSV-1 glycoprotein-mediated cell fusion events. A549 cells were transfected with plasmids expressing the individual HSV-1 glycoproteins (gB, gD, gH, and gL). After 6 h of transfection, cells were washed with PBS and treated with CHLA (60 µM), PUG (40 µM), heparin (100 µg/ml), or DMSO (0.1 %). After further incubation for 24 h, cells were fixed with methanol and stained with Hoescht dye (A) or Giemsa (B). Photomicrographs were then taken at 200× magnification: (A) shows phase (upper panels) and the respective fluorescence pictures displaying the Hoescht dye-stained nuclei (bottom panels); (B) shows the Giemsa-stained cells of similar experiment. Representative pictures shown are from one of three independent experiments. Vector: empty vector; GP: HSV-1 glycoproteins; scale bars: 100 µm. (C) The total number of polykaryocytes (> 10 nuclei) from each treatment was quantified. Data shown are the mean ± SEM of three independent experiments.
FIG. 9. Anti-HSV-1 effects mediated by CHLA and PUG are impaired in GAG-deficient cells and are rescued by restoration of heparan sulfate biosynthesis through EXT1 gene expression. Confluent cells in 12-well plates were infected with HSV-1-GFP (200 PFU/well reflecting viral titers determined in each cell line) concurrently in the presence or absence of CHLA (60 µM), PUG (40 µM), or DMSO (0.1 %) for 1 h. The plates were washed with PBS before applying overlay media. After an additional 48 h of incubation, fluorescent viral plaques were scanned and quantified. Values obtained were compared against each cell line’s respective DMSO control for HSV-1 infection which was considered to be 100 %. Respective status of the heparan sulfate (HS) and chondroitin sulfate (CS) GAG synthesis in the different cell lines are indicated in parentheses. Data shown are the mean ± SEM of three independent experiments with each treatment being performed in duplicate.
<table>
<thead>
<tr>
<th>Compounds</th>
<th>EC50 (µM)</th>
<th>Anti-HSV-1 effects</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
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<tr>
<td>CHL</td>
<td>316.87 ± 0.01</td>
<td>17.02 ± 2.62</td>
</tr>
<tr>
<td>DHE</td>
<td>330.83 ± 9.07</td>
<td>20.05 ± 2.40</td>
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<tr>
<td>PUG</td>
<td>316.84 ± 4.60</td>
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<tr>
<td>PUN</td>
<td>310.85 ± 1.99</td>
<td>21.35 ± 1.77</td>
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<tr>
<td>ACV</td>
<td>&gt; 2000</td>
<td>14.45 ± 1.71</td>
</tr>
<tr>
<td>PGB</td>
<td>&gt; 2000</td>
<td>182.35 ± 20.79</td>
</tr>
</tbody>
</table>

a. Compounds were evaluated by MTT assay to determine the concentration of 50% inhibition of viral yield (EC50)

b. Inhibition was calculated by compare assay to determine the effective percentage reduction of wild type (EC50).

c. IC50 = EC50 / 0.5

d. EC50 = 100 IC50

Values shown are means ± SEM derived from the mean of three independent experiments with each treatment in triplicate.