Therapy with minocycline aggravates experimental rabies in mice

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Abstract

Minocycline is a tetracycline-derivative with anti-apoptotic and anti-inflammatory properties, and the drug has been shown to have beneficial effects in a variety of models of neurological disorders. In this study the potentially neuroprotective role of minocycline was assessed in experimental in vitro and in vivo models of rabies virus infection. In this study 5 nM minocycline did not improve the viability of embryonic mouse cortical and hippocampal neurons infected in vitro with the attenuated SAD-D29 strain of rabies virus (D29) assessed using trypan blue exclusion. Two-day-old ICR mice were inoculated in the right hindlimb thigh muscle with D29 and they received daily subcutaneous injections of either 50 mg/kg minocycline or vehicle (PBS). Infected minocycline-treated mice experienced an earlier onset of neurologic signs and greater mortality (83% vs. 50%) than those receiving vehicle (log rank test, p=0.002 and p=0.003, respectively). Immunohistochemical analysis of rabies virus antigen distribution was performed at early time points and in moribund mice. There were greater numbers of infected neurons in regional brain areas of minocycline-treated mice, which was significant in the CA1 region of the hippocampus. There was less apoptosis (p=0.01) and caspase 3 immunostaining (p=0.0008) in the midbrains of mice treated with minocycline than with vehicle, consistent with a neuroprotective role of neuronal apoptosis that may have had a mild effect inhibiting viral spread. Reduced infiltration of CD3+ T-cells was observed in the pons/medulla of moribund mice that received minocycline therapy (p=0.008), suggesting that the anti-inflammatory actions of minocycline may intensify the neurologic disease. These findings indicate that minocycline has important detrimental effects in the therapy of experimental rabies. Empirical therapy with minocycline should therefore be approached with caution in human rabies and possibly other viral encephalitides until more experimental data become available.
Introduction

New therapeutic agents are needed for the therapy of rabies encephalitis in humans. Despite initial enthusiasm about therapy with the anesthetic agent ketamine (6), recent experimental (19) and clinical (4) evidence have raised questions about the therapeutic usefulness of this agent in human rabies. A combination of therapeutic agents is a reasonable approach for consideration in rabies as used in other viral infections and other diseases (6). Minocycline is a tetracycline-derivative with anti-apoptotic and anti-inflammatory properties, and the drug has been shown to have beneficial effects in a variety of neurological disorders (21). Efficacy of minocycline therapy has also been reported in a variety of experimental models of viral infections in animals, including neuroadapted Sindbis virus infection of mice (2), reovirus infection of neonatal mice (14), and simian immunodeficiency virus infection of pigtailed macaques (23). In this report we evaluate the efficacy of using minocycline therapy in experimental infection of neonatal mice with rabies virus as well as in rabies virus-infected mouse embryonic neurons. We have selected a model using the SAD-D29 strain of rabies virus with peripheral inoculation in neonatal mice (5), that, with adjustment of the viral dose, produces brain infection with only about a 50% mortality rate and facilitates evaluation of either mild beneficial or detrimental effects of the therapy. Preliminary studies in adult mice with highly virulent rabies virus (challenge virus standard-11) had failed to demonstrate a beneficial effect. We have recently characterized this model and shown an encephalitis with prominent lesions in the brainstem and cerebellum (5).
Materials and Methods

Virus

The SAD-D29 strain (D29) of rabies virus was obtained from Teshome Mebatsion (Intervet International, Boxmeer, The Netherlands) (11). D29 was generated by substitution of arginine (R333) of the mature rabies virus glycoprotein with aspartic acid (D333) using recombinant rabies virus L16, which contains the authentic sequence of the SAD-B19 vaccine strain (11). Baby hamster kidney (BHK) cells (C13 clone) grown in Dulbecco’s modified Eagle medium (DMEM) supplemented with 10% fetal bovine serum (FBS) (Invitrogen Life Technologies, Canada) were used for virus propagation.

Preparation of primary neuron cultures

Primary cultures of mouse cortical and hippocampus neurons were prepared as described previously (1). E17 pregnant CD-1 mice were anesthetized with isoflurane (Baxter Corporation, Canada) and fetuses were immediately removed from the uterus and placed in a petri-dish on ice. Under sterile conditions, the fetuses were separated from the embryonic sacs. The fetal cerebral cortices and hippocampi were collected and digested with trypsin (0.1%) and DNAse I (50 µg/mL). The tissues were pooled in sterile Neurobasal medium (Invitrogen Life Technologies, Canada) supplemented with B27 (2%) and penicillin-streptomycin (100 IU/mL), triturated, and filtered through a 37 µm nylon mesh. The filtrate was centrifuged for 10 minutes at 1000 rpm and resuspended in Neurobasal medium at a concentration of $1.0 \times 10^4$ cells/mL. Cells were then plated onto 24 (1 mL) well plates containing 12-mm round coverslips coated with poly-D-lysine.
(50 µg/mL) and incubated in 5% CO₂ atmosphere at 37°C. The cells were cultivated for 4 days prior to viral adsorption.

Determination of viability of infected primary neuron cultures

Neurons plated in 24- (1 mL) well plates (1 ×10⁴ neurons per well) were infected at a multiplicity of infection of 10 focus-forming units (ffu)/neuron of D29. After viral adsorption for 1 hr, the cells were washed once with PBS and fresh Neurobasal medium with or without minocycline (Cat. # M-9511, Sigma-Aldrich, St. Louis, MO) was added immediately with incubation at 37°C. Media and minocycline were not replenished. At 24, 48, and 72 hr post-infection (p.i.) cells were fixed with 4% paraformaldehyde and viability was determined by trypan blue exclusion with phase-contrast microscopy at high power magnification (40× objective). Three separate experiments were performed. Trypan blue exclusion is a cell viability assay based on the ability of the cells to exclude the vital dye trypan blue, and cells in an early stage of apoptosis would be expected to exclude the dye and score as viable cells (10). Non-viable neurons failed to exclude the vital dye and showed trypan blue staining in their cytoplasm.

Viral yield assays

The supernatants of primary neuronal cultures were assayed by fluorescent focus formation on BHK cell monolayers (7, 16).

Animals and inoculations

Two-day-old ICR mice (Charles River Canada, St. Constant, Quebec) were inoculated in the right hindlimb thigh muscle with 100 ffu of D29 in 20 µL. Uninfected controls were inoculated
with vehicle, phosphate-buffered saline (PBS), in the same manner. Minocycline (50 mg/kg/day) diluted in 0.05 – 0.10 mL PBS was administered once daily subcutaneously for a total of 18 treatments per animal (n = 24) on days 0 to 17 post-infection beginning within 6 hours after viral inoculation. Vehicle (0.05 – 0.10 mL PBS) was given to an equal number (n = 24) of D29-infected mice. A repeat (second) experiment was performed with 60 D29-infected mice treated with vehicle or minocycline in order to verify the clinical observations and mortality data. A separate (third) experiment was performed in order to assess the number of infected neurons on days 5 and 7 p.i. in vehicle- (n = 12) and minocycline- (n = 12) treated mice.

Clinical observations

Mice were evaluated daily for 28 days p.i. for clinical neurological signs, including hunching, limb paresis, and ataxia. Moribund animals were euthanized. Deaths occurring during the first 3 days p.i. were excluded because they were likely unrelated to the infection.

Preparation of tissue sections

Mice were anesthetized with isoflurane and perfused with buffered 4% paraformaldehyde between days 3 to 7 p.i., or after they developed clinical rabies and became moribund. Brains were removed and immersion-fixed in the same fixative for 24 h at 4°C. Coronal tissue sections (6 µm) were prepared after dehydration and embedding in paraffin and stained with cresyl violet for light microscopic examination.

Assessment of neuronal apoptosis in mouse tissues

Neuronal apoptosis was graded in major regional brain areas of minocycline- and vehicle-treated mice. A semiquantitative evaluation of the severity of apoptotic changes (40× objective) was
performed with the following scheme: 0, no significant changes; 1, mild changes; 2, moderate changes; 3, severe changes; and 4, very severe changes and/or neuronal loss. The identity of all slides was masked during scoring. Rating scale scores are expressed as the mean score ± standard error of the mean. Treatment with vehicle or minocycline was compared using the Mann-Whitney test.

**Immunohistochemistry**

For expression of rabies virus antigen, cleaved (activated) caspase 3, and MAP-2, primary neurons were grown on poly-D-lysine coated glass coverslips. At various times following D29 infection, neurons were fixed with 4% paraformaldehyde-PBS at 4°C overnight and analyzed. To detect rabies virus antigen, cells were washed in PBS, blocked for 20 minutes in 5% rabbit serum-PBS and then incubated overnight at 4°C with a mouse anti-rabies virus nucleocapsid protein monoclonal antibody (5DF12), which was obtained from A.I. Wandeler (Centre of Expertise for Rabies, Canadian Food Inspection Agency, Nepean, ON, Canada) at 1:160 dilution in 2% rabbit serum-PBS. For caspase 3, cells were washed in PBS, blocked for 20 minutes in 5% goat serum-PBS and then incubated overnight at 4°C with polyclonal rabbit antibody against caspase 3 (Cat. No. 9661, Cell Signaling Technology, Inc., USA) at 1:200 dilution in 2% goat serum-PBS. For identification of the presence of a neuronal marker, cells were washed in PBS, permeabilized in PBS containing 0.5% Triton X-100 for 10 minutes at room temperature, blocked for 1hr in 5% rabbit serum-PBS and then incubated overnight at 4°C with anti-MAP-2 (Sigma-Aldrich, St. Louis, MO) at 1:200 dilution in 5% rabbit serum-PBS. The neurons were washed twice in PBS, incubated for 30 minutes with secondary antibody biotinylated rabbit anti-mouse IgG (for viral antigen and neuronal marker detection) or goat anti-rabbit IgG (caspase 3
activity) IgG (Vector Laboratories Inc., Canada) at 1:100 dilution in 2% normal rabbit or normal goat serum-PBS, respectively. Neurons were washed twice in PBS, incubated for 30 minutes with 1.0% H₂O₂ in methanol. The ABC kit (Vector Laboratories Inc.) was used to localize the biotinylated antibody and diaminobenzidine was used as a substrate for color development. Slides were counterstained with Gill’s hematoxylin and examined using light microscopy.

For rabies virus antigen detection in tissue sections, the sections were deparaffinized and rehydrated prior to incubation with 1.0% H₂O₂ in MeOH. Tissue was subsequently treated overnight at room temperature with mouse anti-rabies virus nucleocapsid protein monoclonal antibody 5DF12 (see above) diluted 1:160, DakoCytomation EnVision+ System-HRP labeled polymer anti-mouse (DakoCytomation, Denmark) for 40 min, 0.5mg/ml 3,3-diaminobenzidine tetrachloride in 0.01% H₂O₂ in PBS, and 0.5%CuSO₄ in 0.15M NaCl. Slides were counterstained with hematoxylin and examined using light microscopy.

Immunohistochemical staining for activated caspase 3 was performed using a polyclonal rabbit antibody against caspase 3 (Cat. No. 9661, Cell Signaling Technology, Inc., USA) as described (13); immunohistochemical staining of inflammatory T cells was done using a CD3 marker (polyclonal anti-human CD3, DakoCytomation, Glostrup, Denmark) diluted 1:400 as primary antibody.

Statistical analyses

Data were analyzed using t-tests for the significance of the difference between the means of two counted samples and Mann Whitney tests for the significance between two independent samples
using a semiquantitative scoring system (Fig. 3A), as well as Kaplan-Meier survival curves and log rank test values of p<0.05 were considered statistically significant.

Results

Characterization of primary neuron cultures

Over 95% of the cells in the cortical and hippocampal cultures expressed the neuronal marker MAP-2 using immunohistochemical staining (data not shown). By 72 hrs p.i. there was immunostaining of rabies virus antigen in 78% of D29-infected cells (data not shown), while staining was not observed in mock-infected cultures. At 24, 48, and 72 hrs p.i., 17%, 26%, and 28% of D29-infected cells expressed activated caspase 3, respectively (data not shown).

Dose-related effects of minocycline on viability of primary neuronal cultures

We initially evaluated viability of cortical neurons with minocycline over a concentration range of 0 – 200 nM. A concentration of 5 nM was selected for further studies because it showed only small reductions in viability (5, 3, and 0% at 24, 48, and 72 hrs p.i., respectively) on screening toxicity studies in cultures (data not shown), and was similar to the concentration used by others using embryonic neurons (9).

D29 infection resulted in loss of viability in primary neuronal cultures

Neurons in the cortical (Fig 1A) and hippocampal (Fig. 1B) cultures were infected with D29 or mock-infected and analyzed for viability using trypan blue exclusion at days 1, 2, and 3 p.i. (Fig. 1A and B). There was a progressive loss in the viability of the cortical and hippocampal cultures over time for the D29-infected neurons compared with the mock-infected cultures.
Minocycline did not exert a neuroprotective effect in D29-infected cultures

In three separate experiments there were small reductions in the viability of cultures with 5 nM minocycline versus cultures without minocycline (Fig. 1A and B). In this system, 5 nM minocycline did not affect the viral yield in cortical cultures (Fig. 1C) and only demonstrated a mild reduction in viral yield at 3 days p.i. in the hippocampal cultures (Fig. 1D).

Clinical observations of mice

Forty-eight two-day-old mice were inoculated in the hindlimb thigh muscle with 100 focus-forming units of D29 and were given either minocycline (50 mg/kg/day) or vehicle (PBS) administered once daily subcutaneously for a total of 18 treatments. Both cumulative clinical neurological disease, including paresis and ataxia, and mortality (83% vs. 50% in vehicle-treated controls) were significantly more marked in the minocycline-treated mice (log rank test, p=0.002 and p=0.003, respectively) (Fig. 2). The larger repeat experiment (with 60 infected mice) confirmed greater cumulative clinical neurological disease and mortality in the minocycline-treated mice (log rank test, p=0.006 and p=0.0002, respectively) (data not shown). Clinical improvement of limb paresis occurred in a minority of mice (27%) in both treatment groups. Hence, minocycline actually aggravated the neurological disease and increased the mortality rate of the rabid mice in this model, rather than providing neuroprotection.

Rabies virus antigen distribution in mouse tissues

We compared the number of neurons expressing rabies virus antigen in the brains of minocycline- and vehicle-treated moribund mice pooled from days 9 – 21 p.i. (Fig. 3). Generally,
we found greater numbers of infected neurons in regional brain areas of the minocycline-treated mice, which reached statistical significance in CA1 pyramidal neurons, indicating only modest evidence of more widespread infection in the brains of minocycline-treated mice. In a separate experiment mice were assessed on day 5, before the onset of clinical neurological disease, and 3 of 8 (38%) minocycline-treated mice and 2 of 8 (25%) vehicle-treated mice were found to have infected neurons in the brain. On day 7, 3 of 4 (75%) minocycline-treated mice and 1 of 4 (25%) vehicle-treated mice (with right hindlimb paresis) had infected neurons in the brain, predominantly in the brainstem and cerebellum.

Neuronal apoptosis in mouse tissues

Minocycline-treated mice showed less neuronal apoptosis in the cerebellum and midbrain than vehicle-treated mice, which achieved statistical significance in the midbrain (Mann-Whitney test, p=0.012) (Fig. 4A).

Immunohistochemical evaluation of activated caspase 3 and CD3 expression in mouse tissues

Immunohistochemical evaluation of the number of neurons expressing activated caspase 3 (13), which is a downstream executioner of the apoptotic process, also showed less expression in minocycline-treated mice, and was highly significant in the midbrain (t test, p= 0.0008) (Fig. 4B). Immunohistochemical evaluation of inflammatory T cells using a CD3 marker (polyclonal anti-human CD3, DakoCytomation, Glostrup, Denmark) showed less CD3-positive cells in minocycline-treated mice than in vehicle-treated mice, with statistical significance in the pons/medulla (t test, p=0.008) (Fig. 4C).
Discussion

Because of its neuroprotective anti-apoptotic and anti-inflammatory properties, minocycline shows promise as a future therapy for a variety of neurological diseases, including multiple sclerosis, spinal cord injury, amyotrophic lateral sclerosis, Parkinson’s disease, and Huntington’s disease (21). A large phase III clinical trial in humans is presently in progress assessing the efficacy of minocycline therapy for amyotrophic lateral sclerosis (17). Some of the potential mechanisms of action of minocycline include anti-apoptotic and anti-inflammatory properties, suppression of free radical production, and inhibition of matrix metalloproteinases (21). Efficacy of minocycline therapy has been reported in a neonatal mouse model of reovirus infection (14). In this model minocycline therapy (35 mg/kg intraperitoneally daily beginning 2 days post-infection) delayed the onset, progression, and mortality of the encephalitis by 4 days, and there was less apoptosis and reductions in viral titer and viral antigen expression in minocycline-treated mice compared with controls. In neuroadapted Sindbis virus infection minocycline (50 mg/kg intraperitoneally daily beginning at start of infection) protected mice from the development of hindlimb weakness and from viral-induced motor neuron death without effects on viral replication or spread (2). In simian immunodeficiency virus infection of pigtailed macaques, minocycline therapy (4 mg/kg/day orally) reduced the severity of the encephalitis, suppressed the viral load in the brain, and decreased the expression of inflammatory markers in the brain (23). An effective neuroprotective agent would be an exciting advance in the therapy of viral encephalitis in humans, and there is a temptation to use the drug empirically with the hope that it might have a beneficial effect before the results of further studies in animals or humans become available. This is especially true in rabies, which is almost always fatal.
There is abundant evidence that rabies virus strains induce apoptosis of infected rodent embryonic neurons in vitro (12, 19). Furthermore, more attenuated rabies virus strains have a greater capacity to induce neuronal apoptosis in vitro (12). We have recently shown that the highly attenuated D29 strain with a mutation at position 333 in the rabies virus glycoprotein also has a greater capacity to induce neuronal apoptosis in vivo than the less attenuated L16 strain from which it was derived. Minocycline did not produce neuroprotection of D29-infected embryonic neurons in the present study. Although D29 infection is associated with increased expression of activated caspase 3 in the embryonic neurons, there may also be D29-induced non-apoptotic mechanisms of cell death and caspase-independent mechanisms of apoptotic cell death, as previously reported in rabies virus infection (15), accounting for the loss of viability in these primary neurons.

In the present neonatal mouse model, minocycline treatment of rabies virus-infected mice was associated with more severe neurological disease and a higher mortality rate than treatment with vehicle. The number of infected neurons in regional brain areas of moribund mice treated with minocycline was greater than in those treated with vehicle, but this only reached statistical significance in the CA1 region of the hippocampus. In mice assessed at earlier time points, including before the onset of clinical neurological disease, there was a trend towards finding more infected neurons in the brains of minocycline- than vehicle-treated mice. This suggests mildly more efficient viral spread in the minocycline-treated mice, which could reflect effects on host defenses, including both anti-inflammatory and anti-apoptotic effects.
Although minocycline did not improve the viability of D29-infected embryonic neurons \textit{in vitro}, the drug exerted an anti-apoptotic effect \textit{in vivo}, similar to the observations of Richardson-Burns and Tyler (14) in neonatal reovirus infection. The neuroprotective effects in the reovirus model may have occurred because neuronal apoptosis plays a more fundamental role in the pathogenesis of the disease, rather than mainly a mechanism for host defense by inhibiting viral spread (5, 12). There were less CD3-positive cells in the brain parenchyma of minocycline-treated D29-infected mice than in vehicle-treated mice, which reached statistical significance in the pons/medulla. This indicates an anti-inflammatory effect of minocycline, which may have also affected other immune effectors and impaired the host’s ability to control the infection, resulting in aggravation of the rabies encephalomyelitis with a higher mortality rate. Both innate and adaptive immunity are important for recovery from rabies virus infection (8), and we feel that the anti-inflammatory effects of minocycline were likely important in aggravating the rabies encephalitis.

Minocycline has also been recognized to aggravate disease in animal models of neurodegenerative diseases. In the 3-nitropropionic acid mouse model of Huntington’s disease, minocycline treatment was associated with a worse mean motor score, impaired general activity, and significantly deteriorated performances on the rotarod, pole test, and beam-traversing tasks and was associated with more severe neuronal cell loss in the dorsal striatum in comparison with untreated mice (3). In minocycline-treated monkeys in a 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) model of Parkinson’s disease, the same investigators also found more severe clinical features of parkinsonism and greater loss of putaminal dopaminergic nerve endings versus untreated animals (3). Minocycline was also found to increase MPTP-induced
damage to dopaminergic neurons in mice (20). Although minocycline has been found to protect against cerebral ischemia in a rat model of transient middle cerebral artery occlusion (22), the opposite effect has been observed in a mouse model of hypoxic-ischemic brain injury (18).

Given the present finding that minocycline therapy aggravated rabies encephalitis in an experimental mouse model, we recommend that minocycline not be administered to human patients with rabies and possibly other viral encephalitides on an empirical basis, because of its potential ability to aggravate the disease. More studies are needed to understand the complex effects of minocycline in different neurological diseases, including rabies and other viral encephalitides.
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References


Figure Legends

Fig. 1. Viability of mock- and D29-infected cultured cortical (A) and hippocampal (B) neurons with and without treatment with 5 nM minocycline as assessed by trypan blue exclusion. Minocycline produced small reductions (4.1 – 7.6% in cortical neurons and 0.7 – 5.2% in hippocampal neurons) in the viability of the D29-infected neurons. Viral yield assays of culture supernatants expressed as mean titers (ffu/mL) from cortical (C) and hippocampal neurons (D). * represents statistical significance with p<0.05 using unpaired t-test and error bars represent the standard error of the mean for comparisons of values for mock infection without minocycline vs. with minocycline and of D29 infection without minocycline vs. with minocycline.

Fig. 2. Cumulative clinical neurological signs and cumulative mortality for vehicle- (n=24) and minocycline- (n=24) treated mice.

Fig. 3. Counts of the number of infected neurons in various brain regions of moribund mice (9-21 days p.i. that received daily treatment with vehicle (n=9) or minocycline (n=12). Slides stained for rabies virus antigen were masked and the numbers of infected neurons were counted in three different fields of the same brain region using a high power (40×) objective in areas with the most marked staining. * represents statistical significance with p<0.05 using unpaired t-test and error bars represent the standard error of the mean for comparisons of values for treatments with vehicle vs. minocycline.

Fig. 4. Morphological changes of apoptosis were evaluated in neurons in the midbrain of moribund mice (9-21 days p.i.) treated with either vehicle (n=9) or minocycline (n=12) daily (A)
as described in the Materials and Methods. Counts of the number of neurons expressing caspase 3 (B) in the midbrain and cerebellum and of CD3-positive cells (C) in the diencephalon, midbrain, and pons/medulla of moribund mice that received daily treatment with vehicle (n=9) or minocycline (n=12). Slides stained for caspase 3 or CD3 were masked and the numbers of stained neurons and CD3-positive cells were counted by two independent observers in three different fields of the same brain region using high power (40×) objective in areas with the most marked staining. * represents statistical significance with p<0.05 using unpaired t-test and error bars represent the standard error of the mean for comparisons of values for treatments with vehicle vs. minocycline.