Optimal Induction of T-Cell Responses against Hepatitis C Virus E2 by Antigen Engineering in DNA Immunization

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Although DNA immunization is a safe and efficient method for inducing cellular immune responses, it generates relatively weak and slow immune responses. Here, we investigated the effect of hepatitis C virus (HCV) antigen modifications on the induction of T-cell responses in DNA immunization. It is likely that the strength of T-cell responses has an inverse relationship with the length of the insert DNA. Interestingly, a mixture of several plasmids carrying each gene induced a higher level of T-cell responses than a single plasmid expressing a long polyprotein. Moreover, the presence of a transmembrane domain in HCV E2 resulted in stronger T-cell responses against E2 protein than its absence. Taken together, our results indicate that the tailored modifications of DNA-encoded antigens are capable of optimizing the induction of T-cell responses which is required for eliminating the cells chronically infected with highly variable viruses such as HCV and human immunodeficiency virus.

Since the first report of DNA immunization in 1990 (45), it has been shown to be efficacious in many disease models. (7, 36, 38). DNA vaccines mimic live attenuated vaccines in their ability to induce major histocompatibility complex (MHC) class I-restricted CD8+ T-cell responses while mitigating some of the safety concerns associated with live vaccines. In particular, DNA vaccines can eliminate the need for a cold chain (17).

However, despite early encouraging results, the level of specific immunity induced by DNA vaccines has generally been regarded as insufficient to offer protection against highly pathogenic organisms. To overcome this obstacle, many approaches have been taken to improve the efficacy of DNA vaccines, such as the incorporation of genes for cytokines and costimulatory molecules, the insertion of additional CpG motifs, in vivo electroporation, and the use of self-replicating viral vectors, in particular, DNA vaccines can eliminate the need for a cold chain (17).

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shorter-length inserts (16). One of the most critical concerns to be addressed is the relationship between insert length in a plasmid and the strength of induced T-cell responses.

Immunological research in the HCV field has been difficult, because there is no reliable small-animal model. A chimpanzee is the only relevant animal model for HCV infection; however, chimpanzees are costly and rare and have to be used judiciously. The limitations of the chimpanzee model have hampered testing of a variety of strategies designed to enhance immunity to HCV antigens. Although the use of mice does not permit HCV replication, studies using a mouse model have contributed to our understanding of the generic aspects of immunization, such as the nature of antigens and the route, time course, and order of delivery of immunogens for the generation of optimal immune responses (including the use of cytokines or other possible immunomodulators). In this study, we investigated the effect of HCV antigen modifications on the optimization of the induction of strong cellular immune responses. HCV E2 was selected as a reporter antigen, because the analysis of immune responses to E2 has been previously established (18). Using different lengths of HCV antigen-expressing DNA constructs, we demonstrated that the T-cell response was inversely correlated with the length of the insert. In addition, immunization with a mixture of separate plasmids which each expressed a protein induced stronger T-cell responses than immunization with a plasmid expressing a long polypeptide. Finally, deletion of the transmembrane domain (TMD) of E2 resulted in a reduction of T-cell responses.

MATERIALS AND METHODS

Animals and immunization. Female BALB/c mice (H-2d) 5 to 6 weeks of age were purchased from Japan SLC (Shizuoka, Japan) and maintained under specific pathogen-free conditions. Mice (five to six per group) were immunized with 100 µg of an indicated plasmid DNA injected bilaterally into the anterior tibialis muscle. A control group of mice were injected with vector DNA alone. At the indicated time after immunization, mice were sacrificed and spleenocytes were removed for the analysis of cell-mediated immune responses. Animals are maintained in accordance with applicable portions of the Animal Welfare Act and with the guidelines of the United States Department of Health and Human Services, Public Health Services, the National Institutes of Health, and the Guide for the Care and Use of Laboratory Animals.

Construction of expression plasmids. pTZ-HCV (encoding a whole genomic region of HCV type 1b, which consists of a structural region from a Korean isolate [accession number AY308072] and a nonstructural region from a JS strain) (8, 41) was generated using restriction enzyme sites within HCV sequences on the basis of a standard molecular cloning technique (39) and used as a template for PCR amplifications in this study. pTV2-sE2t, which localized E2 protein into the endoplasmic reticulum by inserting the signal sequence of herpes simplex virus glycoprotein D into the N terminus of E2, was described previously (27). To construct pTV2-ST, pTV2-SN2, and pTV2-6NST, PCR amplifications using Core PriIS (5′-aag ctc cag acc atg agc aga cca aat cct aat ct-3′), E2 XboIA (5′-ccc tct aga tgg gct gac cag gag aag-3′), Core PriIS_NS2_XboIA (5′-aag ctc cag acc atg agc aga cca aat cct aat ct-3′), and Core PriIS (5′-aag ctc cag acc atg agc aga cca aat cct aat ct-3′) were performed and the product was digested with XbaI for insertion into a pTV2 vector. Core XboIS (5′-aag ctc cag acc atg agc aga cca aat cct aat ct-3′) was amplified by PCR using Core PriIS_NS2_XboIA (5′-aag ctc cag acc atg agc aga cca aat cct aat ct-3′) and was ligated to each other. To construct pTV2-s, a PCR product from pTV2-sE2t was prepared using cytomegalovirus AsX1 (5′-aag ctc cag acc atg aga cca gag gga gga gag gga gga ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt ggt g
expressed the core from which 40 aa at the N terminus were deleted, namely, pTV2-ΔST, pTV2-ΔSN2, and pTV2-sΔST. To investigate the effect of the presence of the E2 TMD on the strength of the cellular immune response, the TMD of E2 was truncated from the pTV2-sE2 and pTV2-sΔST plasmids to construct pTV2-sE2t and pTV2-sΔSTt, respectively.

To identify the expression of the above constructs, COS7 cells were transiently transfected with the indicated plasmids followed by Western blot analysis using anti-E2 monoclonal antibodies (25). A luciferase-expressing DNA was also cotransfected with the constructed plasmids, and transfection efficiency was normalized according to the luciferase activity. Expected E2 glycoproteins were detected at around 65 to 70 kDa. The highest and lowest levels of expression were detected in pTV2-sE2t and pTV2-SN5, respectively (Fig. 1b, lanes 3 and 10). pTV2-ST-related constructs appeared to express intermediate levels of E2 protein (lanes 4 to 7). Expression levels of pTV2-SN2 and pTV2-ΔSN2 were between those of pTV2-ST-related constructs and pTV2-SN5 (lanes 8 and 9). These results suggest that the expression levels have an inverse relationship with the insert length of the plasmid. Truncation of E2 TMD in pTV2-sE2t and pTV2-sΔSTt plasmids was identified by the slight difference in their migration characteristics compared to those of pTV2-sE2 and pTV2-sΔST (Fig. 1b, lanes 2 and 6 versus lanes 3 and 7) and resulted in the secretion of E2 protein (data not shown). Although pTV2-ST appeared to express a higher level of E2 than any other pTV2-ST derivatives, it was due to the spreading over of the pTV2-sE2t band, as confirmed by independent experiments. The pattern of relative expression levels of the above constructs were also confirmed when a C2C12 myoblast cell line was used (data not shown).

The effect of insert length on the strength of E2-specific T-cell responses induced by DNA immunization. To investigate the effect of insert length on the induction of cellular immune responses, different plasmids with inserts of increasing length were used for immunization. T-cell responses were analyzed using an E2-expressing CT26-hghE2t cell line at 5 weeks after immunization. pTV2-sE2 appeared to induce the highest IFN-γ ELISPOT response and 84% of cytolytic activity at an E:T ratio of 30. In contrast, pTV2-ST, pTV2-SN2, and pTV2-SN5 induced 47, 23, and 3% of the IFN-γ ELISPOT number induced by pTV2-sE2 and 50, 18, and 2% of specific lysis at an E:T ratio of 30, respectively (Fig. 2a). These results suggest that there is an inverse relationship between insert length and the strength of T-cell responses.

When plasmids with core deletions were tested for the ability to induce E2-specific T-cell responses, pTV2-ΔST, pTV2-ΔST, and pTV2-ΔSN2 induced 77, 84, and 31% of the IFN-γ ELISPOT number induced by pTV2-sE2 and 79, 68, and 53% of specific lysis at an E:T ratio of 30, respectively (Fig. 2b) and thus revealed a similar inverse relationship between insert length and strength of T-cell responses. Interestingly, it is likely that there is an enhancement of IFN-γ ELISPOT response in the expression of plasmids with core deletions compared to that seen with plasmids with intact core; pTV2-ΔST and pTV2-
FIG. 2. HCV E2-specific T-cell responses inversely correlated with insert lengths of DNA constructs. BALB/c mice were immunized intramuscularly with 100 μg of the indicated expression plasmids. Splenocytes were removed at 5 weeks after immunization and used for CTL and IFN-γ ELISPOT assays with CT26-hghE2t cells for stimulation. Plasmids containing intact core (a) or Δcore (b) were compared for their levels of induction of T-cell responses. Standard deviations are indicated as error bars. Data are representative of the results of one of three independent experiments. Statistical analysis was performed using Student’s t test. P values between different groups of immunization were compared, and those of less than 0.01 were considered significant.

ΔSN2 induced 163 and 134% of the number of IFN-γ secreting cells induced by pTV2-ST (P < 0.01) and pTV2-SN2 (P < 0.1), respectively (Fig. 2a and b, respectively), which might be explained by the elimination of the putative immunosuppressive function of core protein.

Effect of antigen expression on the induction level of T-cell responses. It is likely that there is an overall inverse relationship between expression level and insert size, since the expression levels were decreased overall as the insert became longer (E2 > ST > SN2 > SN5) (Fig. 1b). To investigate whether the magnitude of T-cell responses correlated with expression level, we constructed three different plasmids expressing HCV structural proteins, pGX10-sΔST, pGX101-sΔST, and pGX103-sΔST, which express different levels of E2 protein (Fig. 3a); the relative ratios of the E2 expression levels of pGX10-sΔST, pGX101-sΔST, and pGX103-sΔST were approximately 500:10:1 (Fig. 3b). pGX101-sΔST and pGX103-sΔST induced 103 and 45% of the ELISPOT numbers induced by pGX10 sΔST, respectively, at 5 weeks after the first immunization (Fig. 3c). In addition, pGX10-sΔST, pGX101-sΔST, and pGX103-sΔST showed 77, 66, and 61% of CTL activity at an E:T ratio of 30, respectively. At 3 weeks after boosting, the number of spots induced by pGX101-sΔST and pGX103-sΔST were 67 and 68% of the number induced by pGX10-sΔST. These results indicated that the expression level slightly affects the induction level of T-cell response but is not directly proportional to T-cell responses in DNA vaccination. These results are partially consistent with a recent report that antigen expression does not correlate with immunogenicity (28).

The effect of nonstructural gene coexpression on the induction of E2-specific CD8+ T-cell responses. T-cell responses induced by pTV2-SN5 were significantly lower than those induced by pTV2-se2 or pTV2-sΔST (Fig. 2). It is possible that nonstructural proteins simultaneously expressed with E2 protein in pTV2-SN5 might cause antigenic competition with E2 protein and thus hinder efficient induction of E2-specific CD8+ T-cell responses. To investigate this possibility, we co-immunized pGX10-sΔST with or without pGX10-NS34 and pGX10-NS5 plasmids to mimic the simultaneous expression of nonstructural proteins in pTV2-SN5 and then compared the induction of E2-specific IFN-γ ELISPOT responses (Fig. 2a). Interestingly, codelivery of pGX10-NS34 with pGX10-sΔST rather slightly (albeit statistically insignificantly) enhanced E2-specific IFN-γ ELISPOT numbers (122%) compared to injection of pGX10-sΔST alone (Fig. 2b). In addition, coinjection of pGX10-NS34 plus pGX10-NS5 appeared to further enhance IFN-γ ELISPOT response (139%). Successful induction of T-cell responses toward nonstructural proteins was confirmed by IFN-γ ELISPOT assays using peptide pools encompassing NS3 or NS5A regions (data not shown). These results suggest that coexpression of nonstructural proteins does not inhibit the induction of T-cell response against E2 in DNA immunization and that antigenic competition by nonstructural proteins is unlikely to be the reason for the low immunogenicity of pTV2-SN5.

The effect of TMD on the strength of cellular immune responses induced by DNA immunization. To investigate the effect of the TMD on the induction of T-cell responses, E2-expressing plasmids (pTV2-se2 and pTV2-sΔST) and TMD-truncated E2-expressing plasmids (pTV2-se2t and pTV2-sΔSt) were used for DNA immunization. To exclude the effect of the contribution of any epitope present in the TMD of E2 on the strength of T-cell response, CT26-hghE2t cells or E2 peptide pools that were devoid of TMD were used for stimulation. At 5 weeks after the first immunization, pTV2-se2 and pTV2-sΔST induced 152 and 200% of the number of IFN-γ spots induced by pTV2-se2t (P < 0.001) and pTV2-sΔSt (P < 0.001), an effect which was further enhanced by booster immunization to 250% (P < 0.005) and 256% (P < 0.004), respectively (Fig. 5). These results indicated that truncation of E2 TMD decreased E2-specific T-cell response no matter how E2 protein was expressed alone or as a portion of polyprotein, suggesting that E2 TMD is required for optimal induction of T-cell responses against E2.

DISCUSSION

In this study, we demonstrated that there is an inverse relationship between insert length and T-cell responses induced by the plasmids tested here. It is possible that a low level of antigen expression might have resulted in reduced T-cell responses, since long inserts led to decreased expression of DNA-encoded antigen. Although the expression level of an antigen has been shown to have a direct relationship with humoral responses in DNA immunization (5), its relationship with T-cell response is still controversial. Previous reports suggested a proportional relationship between expression level and the induced cellular immune responses in DNA immunization with a codon-optimized HIV-1 gag gene or a vaccinia
virus infection model (44, 46). In contrast, no correlation was suggested in other reports of studies in which Venezuelan equine encephalitis virus containing HIV-1 gene or HBsAg and HBcAg DNA vaccine was tested (4, 21). Here, we demonstrated that the expression level appeared to affect (but was not directly proportional to) the level of T-cell responses induced by HCV structural gene immunization. Considering that each antigen has a different level of processing efficiency in the generation of immunogenic peptides which also have different binding affinities to MHC molecules, the nature of the antigen may be another determinant affecting the induction of T-cell responses. For example, when an antigen is efficiently processed into immunogenic peptides that bind to an MHC molecule with high affinity, a small amount of antigen is sufficient to induce a cellular immune response. Furthermore, it is known that the threshold for the induction of T-cell response is lower than that for antibody response (21). Thus, it is likely that the critical factor for inducing T-cell responses in DNA immunization is the nature of the antigen rather than the expression level.

Previous reports demonstrated that there is evidence of antigenic competition in infections with viruses such as lymphocytic choriomeningitis virus and influenza virus (41–43). However, in this study we showed that codelivery of separate plasmids expressing nonstructural proteins of HCV did not affect the induction of E2-specific T-cell response, indicating the absence of competition with coexpressed antigens. The level of antigen expression in DNA immunization in vivo is known to be far less than that in virus infection. This low level of expression may not reach to antigenic competition among dominant and/or subdominant epitopes if target cells provide a number of MHC molecules sufficient to accommodate all of the epitope peptides. Thus, a DNA immunization method may be basically different from virus infection regarding antigenic competition. Moreover, codelivery of plasmids expressing nonstructural proteins rather increased E2-specific T-cell response, presumably due to the presence of T-helper epitopes in the nonstructural region of HCV. It is worth noting that the immunization with a mixture of three DNA constructs (pTV2-ΔST, pTV2-NS34, and pTV2-NS5) was much more efficient at inducing E2-specific ELISPOT activity than immunization with pTV2-NS5 expressing whole HCV polyproteins (>26-fold). These results suggest that antigen processing and presentation induced by a single long polyprotein is inefficient compared to that induced by three separate short polyproteins. It remains to be determined whether the lack of antigenic competition for E2 is applicable to any other proteins within the polyprotein.

It is known that deletion of TMD usually increases the humoral responses by inducing secretion of antigen to the extracellular space, which facilitates antigen encounter with
antigen-specific B cells (12, 39). Although this concept is generally accepted for humoral response, its influence on T-cell response is still controversial. While one group showed that the membrane-bound form of glycoprotein D from bovine herpesvirus induced a stronger CTL response than its secreted form in DNA immunization (29), another group reported there was no difference between the membrane-bound form and the secreted form of ovalbumin in DNA immunization (3). It was of interest to determine how the truncation of E2 TMD decreased IFN-γ ELISPOT responses for E2. Depending on the presence or absence of TMD, a major portion of E2 protein is either cell associated or secreted into the extracellular space (8, 26). One explanation may be the different efficiencies of E2 and TMD-truncated E2 proteins (E2t) in cross-presentation. Dendritic cells were known to process exogenous antigen to load on MHC class I molecules (cross-presentation) and thereby to stimulate CD8+ T cells (20). Several previous reports suggest that cross-presentation is a predominant mechanism for inducing CD8+ T-cell response in DNA immunization (6, 10, 11). During this process, the efficiency of cross-presentation depends on the form of antigen. The cell-associated form of antigen can be cross-presented 103 times more efficiently than the soluble form (30). Thus, the form of antigen provided by a transfected cell could be an important parameter affecting the induction of T-cell response. Also, the induction level of a T-cell response may be influenced by the period of retention time inside the cell before secretion. Thus, it would be disadvantageous for the induction of T-cell responses if secretion were to occur immediately after translation because the shorter retention time within the transfected cells would provide less chance for cross-presentation by dendritic cells.

The balance between T-cell responses and viral replication in HCV infection probably determines whether the infection will result in chronic progression or acute clearance. Thus, optimal induction of T-cell responses through customized modification of an antigen is likely to be necessary for eliminating virus infection. The present study provides information useful for the induction of strong multiemiptope specific T-cell responses by tailored modification of HCV DNA in DNA immunization. Our results can be directly applied to the development of vaccine against HCV and other highly variable infectious agents, such as other RNA and retroviruses which contain polyprotein.

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