STP and Tip Are Essential for Herpesvirus Saimiri Oncogenicity

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Received 22 September 1997/Accepted 27 October 1997

Herpesvirus saimiri (HVS), a gamma-2 herpesvirus or rhadinovirus, infects most squirrel monkeys without apparent disease (9, 13). In other nonhuman primates, however, HVS induces rapidly fatal T-cell lymphoproliferative diseases (14, 17). Sequence divergence among HVS isolates is most extensive at the left end of the unique L-DNA of the viral genome and is the basis for classification of HVS into subgroups A, B, and C (7, 9, 30). Variation in this region is correlated with differences in the capacity of the viruses to immortalize T lymphocytes in vitro and to produce lymphoma in nonhuman primates (4, 7, 8, 10, 24, 33). Viruses of both subgroups A and C immortalize common marmoset T lymphocytes to interleukin-2-independent proliferation (10, 34). Highly oncogenic subgroup C strains also immortalize human, rabbit, and rhesus monkey lymphocytes and can produce fulminant lymphoma in rhesus monkeys as well as in New World primates (1–4, 6, 31).

HVS subgroup A strain 11 mutants with deletions in the first open reading frame at the left end of the genome are capable of replication but fail to immortalize common marmoset T lymphocytes in vitro and do not induce lymphoma in vivo (7, 8, 10, 24, 33). This open reading frame encodes the saimiri transformation-associated protein (STP) (22). HVS subgroup C strain 488 (HVS C488) contains a divergent form of the STP gene along with an additional apparently unrelated open reading frame in the leftmost position (4, 16, 22). Similarities between STPs of HVS subgroup A strain 11 (STP-A11) and STP-C488 and Tip are not required for replication or persistence, but each is essential for transformation in cell culture and for lymphoma induction in common marmosets.

Mutant forms of herpesvirus saimiri (HVS) subgroup C strain 488 with deletions in either STP-C488 or Tip were constructed. The transforming potentials of the HVS mutants were tested in cell culture and in common marmosets. Parental HVS subgroup C strain 488 immortalized common marmoset T lymphocytes in vitro to interleukin-2-independent growth, but neither of the deletion mutants produced such growth transformation. Wild-type HVS produced fatal lymphoma within 19 to 20 days of experimental infection of common marmosets, while HVS ΔSTP-C488 and HVS ΔTip were nononcogenic. Virus was repeatedly isolated from the peripheral blood of marmosets infected with mutant virus for more than 5 months. These results demonstrate that STP-C488 and Tip are not required for replication or persistence, but each is essential for transformation in cell culture and for lymphoma induction in common marmosets.

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MATERIALS AND METHODS

Cell culture and virus propagation. Owl monkey kidney cells (OMK 637) that were cultivated in minimal essential medium supplemented with penicillin, streptomycin, i-glutamine, and 10% (vol/vol) heat-inactivated fetal bovine serum (GIBCO BRL, Grand Island, N.Y.) were used for propagation of HVS C488. Low (<30)-passage OMK cells were used for transfections. Culture of common marmoset lymphocytes in immortalization assays with HVS mutants was performed in RPMI 1640 medium supplemented with penicillin, streptomycin, amphotericin B (Fungizone), i-glutamine, 20% (vol/vol) heat-inactivated fetal bovine serum, and 5 mg of β-mercaptoethanol per liter. COS-1 cells were cultured in Dulbecco modified Eagle medium supplemented with 10% (vol/vol) heat-inactivated fetal calf serum.

Virion DNA isolation. HVS virion preparations were obtained from OMK cell lysates by removal of cell debris by low-speed centrifugation followed by pelleting of the virus at 18,000 rpm for 2 h in an SS-34 rotor. To purify intact virion DNA, the virus was disrupted at 60°C for 2 h in lysis buffer containing 10 mM Tris (pH 8.5), 1 mM EDTA, 1% (vol/vol) Sarkosyl, and 0.1 mg of protease K per ml.
Extraction of the aqueous solution first with an equal volume of phenol and then twice with chloroform was sufficient to purify the virion DNA for use in transformation. Sterile cut pipette tips were used for manipulating virion DNA without shearing.

**Reporter expression plasmid and SEAP assay.** As described previously (11), a reporter gene expression cassette containing the secreted engineered alkaline phosphatase (SEAP) gene under the control of the simian virus 40 (SV40) early promoter and enhancer (SV40-SEAP) was used as a selection marker for the identification of viral recombinants. SEAP production was detected by liquid chemiluminescence assay (Tropix) performed in opaque 96-well microtiter plates following the infection of ligated virion DNA into OMK cells to produce SEAP-positive virus. Transfection of the SEAP expression cassette directly from the recombinant virion DNA was also demonstrated by transforming OMK cells with the reporter gene expression cassette in the absence of virion DNA. Transfection of parental virion DNA into OMK cells produced SEAP-negative virus, while a 42-bp deletion containing more than half of the tip gene was replaced in the Δtip virus (Fig. 1). Recombinants expressing SEAP were isolated by repeated limiting dilution passage in OMK cells.

**RESULTS**

**Isolation of STP-C488 and Tip deletion mutants.** HVS recombinants containing deletions in STP-C488 or Tip were generated by replacing selected portions of a 3.6-kb cloned HVS DNA fragment with a SEAP expression cassette driven by the SV40 early promoter. A 508-bp SpeI/EcoRV fragment including 91% of the STP-C488 gene was replaced with the SEAP expression cassette in the ΔSTP-C488 recombinant, while a 442-bp deletion containing more than half of the tip gene was replaced in the Δtip virus (Fig. 1). Recombinants expressing SEAP were isolated by repeated limiting dilution passage in OMK cells.

To eliminate any possible inadvertent influence of the reporter in transformation assays, AsCl restriction enzyme sites flanking the SEAP expression cassette were used to remove the reporter directly from the recombinant virion DNA. Transformation of ligated virion DNA into OMK cells produced SEAP-negative HVS ΔSTP and Δtip recombinants with the specific gene deletions, which were again purified by repeated passage of limiting dilution of virus. Marker rescued viruses were also generated by recombination between virion DNA from the SEAP-expressing deletion mutants and the 3.6 kb of left-end L-DNA containing authentic STP-C488 and tip genes. Marker-rescued viruses were isolated by screening for SEAP-negative virus and confirming restoration of the deleted regions by PCR and DNA sequencing.

After construction of HVS deletion recombinants, STP-C488 expression was examined by immunoblot analysis. OMK cells were infected with each HVS deletion recombinant, and cell lysates were used for immunoblots with anti-STP-C488 antibody (16). The results revealed that STP was not expressed detectably in cells infected with the STP deletion mutants but was detected in HVS wild type (wt) and ΔTip/SV40-SEAP (Fig. 2A). When the same approach was used to examine Tip expression in deletion viruses, however, we were unable to detect the Tip expression with any of the viruses tested, including wt virus. This was likely caused by the low expression of Tip in HVS.

To demonstrate expression of Tip, we transfected COS-1 cells with the deletion plasmid constructs derived from a 3.6-kb cloned HVS DNA fragment (C488PX) containing wt STP and Tip (10). The C488PX, C488PXΔSTP, C488PXΔSTP/SV40-SEAP, and C488PXΔTip/SV40-SEAP constructs were cotransfected into COS-1 cells together with an Lck expression plasmid. At 48 h after transfection, cell lysates were used for experimental infection of common marmosets. In vivo oncogeticity of the HVS C488 recombinants was assessed by experimental infection of common marmosets. Marmosets were injected intramuscularly with 10⁴ 50% tissue culture infective doses of virus in a volume of 1 ml. Sera and blood cell pellets were collected and frozen at −70°C during the first 4 weeks postinfection and thereafter. Viral loads in PBMC specimens were assessed periodically by duplicate plating of 10⁴ PBMC and serial threefold dilutions of PBMC on OMK cells in 24-well tissue culture plates. Selected culture supernatants from the PBMC viral load plates and selected sera were also tested for SEAP expression from recombinant HVS. Animals that became moribund were euthanized and received complete necropsies. Tissues were fixed in 10% neutral buffered formalin, embedded in paraffin, sectioned, and stained with hematoxylin and eosin.

**In vitro immortalization of common marmoset lymphocytes.** Assays of lymphocyte immortalization in vitro have been described previously (10). Peripheral blood mononuclear cells (PBMC) were isolated from 3-ml heparinized blood specimens from common marmosets (Callithrix jacchus) by centrifugation through lymphocyte separation medium (Organon Teknika Corp., Malvern, Pa.) followed by washing in RPMI 1640 culture medium. PBMC from each animal were individually washed, resuspended in RPMI 1640, and then distributed in 1-ml volumes containing approximately 10⁶ cells into 12-well tissue culture plates. Cells were then infected at a multiplicity of infection ranging from 1 to 5 with 1 ml of purified HVS stocks. Cells were maintained with RPMI 1640 growth medium for 3 to 4 days. Immortalization or cell death was assessed microscopically.

**Experimental infection of common marmosets.** In vivo oncogeticity of the HVS C488 recombinants was assessed by experimental infection of common marmosets. Marmosets were injected intramuscularly with 10⁴ 50% tissue culture infective doses of virus in a volume of 1 ml. Sera and blood cell pellets were collected and frozen at −70°C during the first 4 weeks postinfection and thereafter. Viral loads in PBMC specimens were assessed periodically by duplicate plating of 10⁴ PBMC and serial threefold dilutions of PBMC on OMK cells in 24-well tissue culture plates. Selected culture supernatants from the PBMC viral load plates and selected sera were also tested for SEAP expression from recombinant HVS. Animals that became moribund were euthanized and received complete necropsies. Tissues were fixed in 10% neutral buffered formalin, embedded in paraffin, sectioned, and stained with hematoxylin and eosin.

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immunoprecipitation with anti-Tip antibody, followed by in vitro kinase reaction. The results revealed that Tip was not expressed in C488PX and truncated form of Tip from the C488PX truncated transcriptional unit (Fig. 2). Also, expression of the SEAP reporter expression cassette at the STP locus appeared as efficiently as the wt HVS (Fig. 2B). In fact, repeated experiments contrasted with the wt C488PX (Fig. 2A). In contrast, neither deletion of Tip nor insertion of the reporter gene demonstrated that each of these genes was essential for induction of lymphoma in vivo. The two marmosets infected with wt HVS C488 died on days 19 and 20 after inoculation. However, animals infected with the STP-C488 and/or Tip-deleted recombinants (both with and without the SEAP reporter) remained healthy for the 12 months of observation. Isolation of virus from PBMC of all animals at 3 weeks and beyond verified that all animals were infected. Thus, the STP-C488 and/or tip genes are required not only for T-lymphocyte immortalization in vitro but also for lymphoma induction in vivo.

### In vivo lymphoma induction
Experimental infections of common marmosets with virus deleted of either the STP-C488 or tip gene resulted in loss of the ability to transform common marmoset T lymphocytes in vitro.

To confirm that the loss of transforming activity of deletion virus was derived from the specific gene deletion and not from other unexpected alterations, the STP-C488 or tip gene was restored by homologous recombination of the respective deletion virus with a linearized cloned viral DNA fragment containing the corresponding wt gene. Restoration of STP-C488 or tip genes fully reconstituted the transforming ability of these viruses. Both HVS recombinants with a restored STP-C488 gene or a restored tip gene transformed primary T lymphocytes as efficiently as the wt HVS (Table 1). In addition, we examined the expression of restored STP-C488 by immunoblot analysis using OMK cells infected with the restored HVS. STP was detected at the same level in restored HVS as in the wt HVS C488 (Fig. 2A, lane 5). Also, Tip expression was examined in common marmoset T cells transformed with Tip-restored HVS by in vitro kinase reaction of anti-Tip immune complexes. The 40-kDa phosphorylated Tip along with phosphorylated 56-kDa cellular Lck were detected equally in HVS C488-transformed cells and Tip-restored HVS-transformed cells (Fig. 2C). Thus, the loss of transforming activity in the deletion mutant viruses was attributed solely to the specific STP-C488 or tip gene deletion and not to any inadvertently selected alterations.

### Persistent infection by attenuated HVS deletion viruses in vivo
Antibody responses against HVS were measured by ELISA using plates coated with purified HVS virion proteins. The four marmosets infected with the HVS ΔSTP/SV40-SEAP or HVS ΔTip/SV40-SEAP recombinant showed strong antibody responses to HVS structural antigens. These antibodies persisted at high levels for the entire period of measurement, 20 weeks of infection (Fig. 3).

We have also developed procedures for evaluating virus load in vivo by measuring the numbers of PBMC required to isolate HVS. A similar method has been described previously for

### Table 2

<table>
<thead>
<tr>
<th>Virus*</th>
<th>No. of lymphomas induced/infected marmosets</th>
<th>Survival (days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HVS C488 (wt)</td>
<td>2/2</td>
<td>19–20</td>
</tr>
<tr>
<td>HVS ΔTip</td>
<td>0/2</td>
<td>&gt;370</td>
</tr>
<tr>
<td>HVS ΔTip/SV40-SEAP</td>
<td>0/2</td>
<td>&gt;370</td>
</tr>
<tr>
<td>HVS ΔSTP</td>
<td>0/2</td>
<td>&gt;370</td>
</tr>
<tr>
<td>HVS ΔSTP/SV40-SEAP</td>
<td>0/2</td>
<td>&gt;370</td>
</tr>
</tbody>
</table>

* In all cases, virus was recovered from both marmosoms infected.

### Table 1

<table>
<thead>
<tr>
<th>Virus</th>
<th>No. immortalized/ no. of attempts</th>
</tr>
</thead>
<tbody>
<tr>
<td>None</td>
<td>0/19</td>
</tr>
<tr>
<td>HVS C488</td>
<td>0/19</td>
</tr>
<tr>
<td>HVS ΔTip</td>
<td>23/24</td>
</tr>
<tr>
<td>HVS Tip</td>
<td>0/9</td>
</tr>
<tr>
<td>HVS ΔTip/SV40-SEAP</td>
<td>15/15</td>
</tr>
<tr>
<td>HVS (Tip restored)</td>
<td>0/14</td>
</tr>
<tr>
<td>HVS ΔSTP</td>
<td>12/12</td>
</tr>
</tbody>
</table>
quantitating virus load of simian immunodeficiency virus (23). This assay measures the numbers of infectious cells in PBMC. Virus was readily recovered from PBMC of the animals infected with the deletion viruses for at least 5 months (Fig. 4). While both viruses were readily recovered on repeated occasions, the numbers of infectious cells were consistently higher with the ΔTip virus than the ΔSTP virus whether or not the SEAP gene was present (Fig. 4). Additionally, wt virus-infected marmosets showed a higher level of virus load at week 2 than deletion virus-infected marmosets (Fig. 4).

Assay of SEAP in culture supernatant samples from the viral load determinations at week 21 showed close correlation between detection of SEAP and detection of virus (data not shown). Production of virus capable of SEAP expression through week 21 further indicates persistence of the attenuated viruses in vivo and stability of the SV40-SEAP expression cassette during multiple viral replication cycles in vivo.

Plasma from animals infected with HVS ΔSTP/SV40-SEAP or HVS ΔTip/SV40-SEAP was used to measure the SEAP activity directly. SEAP production was sufficiently strong in the initial weeks after infection to be detected through week 4 (Table 3). Animals infected with the ΔTip/SV40-SEAP virus had consistently higher levels of SEAP activity in plasma during this period than animals infected with ΔSTP/SV40-SEAP (Table 3), consistent with the cell-associated viral load measured as described above.

**DISCUSSION**

The results described here demonstrate that the STP-C488 and tip genes of HVS are required for T-lymphocyte transformation in vitro and lymphoma induction in vivo. Our ability to isolate recombinant viruses with deletions in each of the genes in vitro and to demonstrate persistence in vivo indicates that both genes are nonessential for replication but necessary for transforming capacity. These data are consistent with previous studies demonstrating that sequences nonessential for replication but necessary for transforming capacity are present at the left end of HVS subgroup A genomes (7, 8, 10, 33). However, subgroup A and subgroup C HVS are distinguished by marked divergence in the composition of the leftmost open reading frames and by differences in ability to transform T cells from different species (4). Subgroup C viruses, unlike those of subgroup A, are capable of immortalizing human, rhesus monkey, and rabbit T lymphocytes (1, 3, 29).

Both STP-A11 and STP-C488 are sufficient to transform rodent fibroblasts in cell culture assays (16, 22) and are oncogenic in transgenic mice (25, 32). The extensive sequence divergence of these genes (4, 16, 22) and interaction of their products with distinct cellular targets (19, 26) emphasize the need to independently assess their contributions to oncogenesis through in vivo studies. STP-A11, which interacts with cellular Src, was previously shown to be essential for oncogenesis (33). While the corresponding gene from a different strain of HVS subgroup C has been shown to be necessary for short-term proliferation of T cells (28), STP-C488, which has been shown to associate with cellular Ras (19), has not been assessed previously. The lack of oncogenicity of ΔSTP-C488 deletion virus along with the transforming activity of STP-C488...
TABLE 3. SEAP detection in sera from common marmosets infected with HVS recombinants

<table>
<thead>
<tr>
<th>Wk</th>
<th>Cj1-95a</th>
<th>Cj2-95b</th>
<th>Cj147-91c</th>
<th>Cj362-94c</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1,205</td>
<td>276</td>
<td>0</td>
<td>31</td>
</tr>
<tr>
<td>2</td>
<td>1,634</td>
<td>667</td>
<td>4</td>
<td>18</td>
</tr>
<tr>
<td>3</td>
<td>6,307</td>
<td>1,036</td>
<td>254</td>
<td>0</td>
</tr>
<tr>
<td>4</td>
<td>3,322</td>
<td>1,377</td>
<td>262</td>
<td>0</td>
</tr>
<tr>
<td>6</td>
<td>0</td>
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<td>0</td>
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<tr>
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<tr>
<td>12</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>16</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

a Expressed in chemiluminescent units; average of triplicate measurements.

b Common marmoset infected with HVS ΔSTP/STV40-SEAP.

demonstrates that STP-C488 is necessary for HVS C488 transformation of marmoset T lymphocytes.

Previously, a divergent form of Tip from HVS subgroup C strain 488 has been reported to be required for in vitro T-cell proliferation induced by HVS (27). However, this earlier report did not examine the transforming activity of HVS STP immunization of primary T cells and in lymphoma induction in New World primates. The data reported here show that deletion of Tip in HVS C488 renders the recombinant virus capable of transformation in vitro and in vivo. The role of association between Tip and Lck has been controversial, with both activation (27, 35) and downregulation (15, 21) of Lck-mediated signal transduction being reported. Analysis of HVS with point mutations in the SH3-binding motif of Tip has recently demonstrated that interaction of Tip with Lck is not required for in vitro and in vivo transforming activity of HVS (12). A novel cellular partner of Tip, Tap, which interacts with Tip independently of the Tip-Lck association has recently been described (36). The interaction of Tip with Tap dramatically upregulates NF-κB transcriptional activity and induces the surface expression of lymphocyte adhesion molecules. This finding suggests that study of the interaction of Tip with Tap will be particularly important in defining roles of Tip in HVS oncogenesis.

In addition to insight into the roles of STP-C488 and Tip for viral oncogenesis, we report here new procedures to assess quantitatively the persistence in vivo of attenuated HSVs deletion mutants. While the requirement of STP-C488 and Tip for transforming activity of HVS is now well documented, further detailed study will be required to define the mechanisms utilized by STP and Tip for achieving HVS transformation and for maintenance of high numbers of infected T cells in vivo in natural or nonnatural hosts.

ACKNOWLEDGMENTS

We thank J. Newton, T. Connors, and A. Hampson for manuscript preparation.

This work was supported by Public Health Service grants CA31363 and AI38131 and grant RR00168 from the Division of Research Resources.

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