Passive Protection against Rotavirus-Induced Diarrhea by Monoclonal Antibodies to Surface Proteins vp3 and vp7

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Monoclonal antibodies directed against two rotavirus surface proteins (vp3 and vp7) as well as a rotavirus inner capsid protein (vp6) were tested for their ability to protect suckling mice against virulent rotavirus challenge. Monoclonal antibodies to two distinct epitopes of vp7 of simian rotavirus strain RRV neutralized RRV in vitro and passively protected suckling mice against RRV challenge. A monoclonal antibody directed against vp3 of porcine rotavirus strain OSU neutralized three distinct serotypes in vitro (OSU, RRV, and UK) and passively protected suckling mice against OSU, RRV, and UK virus-induced diarrhea. The role of vp3 in eliciting protection against heterotypic rotavirus challenge should be considered when developing a vaccine with cloned rotavirus genes. Alternatively, immunization with a reassortant rotavirus containing vp3 and vp7 from two antigenically distinct rotavirus parents might protect against diarrhea induced by two or more rotavirus serotypes.

Rotaviruses are an important cause of acute gastroenteritis in infants and young children in developed countries (6, 16). In less developed countries, rotavirus infections often result in diarrhea, dehydration, and death (3, 33).

Parenteral immunization of animals with rotavirus strains heterotypic to a challenge virus has been found to protect against rotavirus-induced gastroenteritis (20, 27, 36–38). Large-scale clinical trials with human infants have demonstrated protection against illness with virulent human strains by oral immunization with a heterotypic bovine rotavirus (30, 31). Litt is known about the molecular basis of protection against rotavirus-induced diarrhea evoked by immunization with strains homotypic or heterotypic to the challenge virus. This information may be relevant to the development of a successful live virus vaccine or a vaccine which utilizes cloned rotavirus genes.

To determine which rotavirus proteins stimulate protective immunity in vivo, we evaluated monoclonal antibodies directed against two rotavirus surface proteins (vp3 and vp7) as well as an inner capsid protein (vp6) for their ability to protect suckling mice against virulent rotavirus challenge (Table 1). Genetic studies using reassortant rotaviruses have found that only gene segments which code for vp3 and vp7 cosegregate with neutralization specificities (15, 18). Monoclonal antibodies were administered orally, since high titers of circulating rotavirus-specific antibodies fail to protect suckling mice against rotavirus challenge and resistance to clinical illness is best correlated with the presence of rotavirus-specific antibodies at the intestinal cell surface (19). Also, oral administration of monoclonal antibodies closely approximates the passive transfer of colostrum and milk antibodies found in breast-fed neonates.

Studies were performed by using a previously developed murine model for oral infection with a tissue culture-adapted primate rotavirus (simian strain SA11) (21). We recently found that suckling mice orally inoculated with the simian rotavirus strain RRV, the porcine strain OSU, or the bovine strain UK developed gastroenteritis similar to that found after oral inoculation with SA11 virus (data not shown) (12).

Five-day-old CD-1 mice were orally inoculated with 100 μl of various dilutions of monoclonal antibodies derived from ascitic fluids. Serum obtained from a BALB/c mouse before inoculation with hybridoma cells was used as a control. After 30 min, suckling mice were orally challenged with 6 × 106 PFU of RRV, SA11, or OSU rotavirus or with 107 PFU of UK rotavirus; these doses caused disease in approximately 95% of animals previously inoculated with control serum (Table 1).

Rotaviruses of six well-characterized serotypes, human strains Wa (serotype 1), S2 (serotype 2), and Saint Thomas (serotype 4), as well as the porcine strain OSU (serotype 5), the simian strains SA11 and RRV, and the bovine strain UK (serotype 6) were used in these studies (14). The simian strains SA11 and RRV have been found to be indistinguishable from a serotype 3 human strain by plaque reduction neutralization (PRN) (14).

Production of monoclonal antibodies by immunization of BALB/c mice with RRV (simian), OSU (porcine), and Wa (human) rotaviruses, fusion of spleen and NS-1 cells, screening of culture supernatant fluids by radioimmunooassay, and production of ascitic fluids in seronegative BALB/c mice were performed as previously described (10, 11). All monoclonal antibodies were isotype immunoglobulin G1, determined by using 125I-labeled goat anti-mouse immunoglobulin G1 (Cappel Laboratories, Malvern, Pa.). Three monoclonal antibodies (2G4, 8B3, and 7A12) which immunoprecipitated vp3 from OSU- and RRV-infected cell lysates (data not shown) were studied for their ability to protect suckling mice against challenge (Table 1). Antibody 2G4 was isolated by fusing lymphocytes obtained from mice parenterally immunized with the porcine OSU strain. Monoclonal antibody 2G4 neutralized the serotypically distinct OSU, serotype 3 (SA11, RRV), and UK strains by PRN and protected suckling mice against diarrhea induced by OSU, RRV, SA11, or UK virus challenge (Table 1). The neutralizing activity of monoclonal antibody 2G4 in vitro against RRV and SA11 was greater than that against OSU. Our finding of a monoclonal antibody (2G4) with neutralizing activity against a heterotypic strain greater than that against the immunizing virus has been described in other systems (34, 35). Mono-
TABLE 1. Protection against RRV-, SA11, or OSU-induced diarrhea in suckling mice by monoclonal antibodies derived from ascitic fluids

<table>
<thead>
<tr>
<th>Monoclonal antibody designation</th>
<th>Immunizing antigen</th>
<th>Structural specificity</th>
<th>RIAa</th>
<th>PRNb titer against the following viruses:</th>
<th>In vivo protective titerc against the following viruses:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Wa</td>
<td>S2</td>
</tr>
<tr>
<td>2G4</td>
<td>OSU</td>
<td>vp3</td>
<td>1.4</td>
<td>10^6</td>
<td>&lt;50</td>
</tr>
<tr>
<td>8B3</td>
<td>RRV</td>
<td>vp3</td>
<td>4.0</td>
<td>10^5</td>
<td>&lt;50</td>
</tr>
<tr>
<td>7A12</td>
<td>RRV</td>
<td>vp3</td>
<td>4.0</td>
<td>10^5</td>
<td>&lt;50</td>
</tr>
<tr>
<td>255/60</td>
<td>RRV</td>
<td>vp6</td>
<td>2.0</td>
<td>10^3</td>
<td>&lt;50</td>
</tr>
<tr>
<td>159-H11</td>
<td>RRV</td>
<td>vp7</td>
<td>1.2</td>
<td>10^5</td>
<td>&lt;50</td>
</tr>
<tr>
<td>4F8</td>
<td>RRV</td>
<td>vp7</td>
<td>1.0</td>
<td>10^6</td>
<td>&lt;50</td>
</tr>
<tr>
<td>4C3</td>
<td>RRV</td>
<td>vp7</td>
<td>1.2</td>
<td>10^5</td>
<td>&lt;50</td>
</tr>
<tr>
<td>5H3</td>
<td>RRV</td>
<td>vp7</td>
<td>1.0</td>
<td>10^6</td>
<td>&lt;50</td>
</tr>
<tr>
<td>2C9</td>
<td>WA</td>
<td>vp7</td>
<td>3.0</td>
<td>10^5</td>
<td>31,250</td>
</tr>
<tr>
<td>129-B8</td>
<td>WA</td>
<td>vp7</td>
<td>1.0</td>
<td>10^5</td>
<td>&lt;50</td>
</tr>
<tr>
<td>6F2-B2</td>
<td>WA</td>
<td>vp7</td>
<td>1.0</td>
<td>10^6</td>
<td>&lt;50</td>
</tr>
<tr>
<td>Serum control</td>
<td></td>
<td></td>
<td>&lt;1.0</td>
<td>10^5</td>
<td>&lt;50</td>
</tr>
</tbody>
</table>

a Conventionally bred BALB/c and pregnant CD-1 mice obtained from Charles River Breeding Laboratories, Inc. (Wilmington, Mass.) were housed in isolation units. Mice were bled on arrival by capillary plexus puncture, and sera were tested by RIA using SA11 virus as previously described (22). Sera obtained from newborns or sera from seronegative CD-1 dams were used in these studies. Viral growth, purification, quantitation, and infectivity titration by plaque assay were performed as previously described (23).

b For a determination of the structural specificities of monoclonal antibodies, [1^53]methionine-labeled RRV-infected cell lysates and radioimmunoprecipitation of labeled proteins with hybridoma culture fluid supernatants were performed as previously described (11). Monoclones 2C9, 4F8, and 4C3 did not immunoprecipitate vp7 efficiently. The structural specificities of these monoclonal antibodies were demonstrated by hemagglutination inhibition using reassortant rotaviruses or by viral variant analysis as previously described (11).

c The radioimmunoblot assay. The radioimmunoblot assay using either RRV or Wa (2C9) as the detecting antigens was performed as previously described (22). Titters are expressed as the reciprocal of the positive ascitic fluid dilution.

* The PRN assay was a modification of the technique described by Matsuno et al. (17), and was performed as previously described (22). Titters are expressed as the reciprocal of the ascitic fluid dilution showing a 50% reduction in the mean plaque count for each virus. (22).

* The reciprocal of the dilution of the ascitic fluid at which 50% of newborn mice were protected against RRV, SA11, OSU, or UK rotavirus challenge.

Monoclonal antibodies 8B3 and 7A12, isolated from mice immunized with RRV, efficiently neutralized RRV in vitro, but failed to protect suckling mice against RRV challenge; the basis for the failure of these antibodies to protect in vivo awaits further study. Competition antibody binding assays and viral variant analysis indicated that monoclonal antibody 2G4 recognized an epitope distinct from that recognized by monoclonal antibodies 8B3 and 7A12 (R. Shaw and H. B. Greenberg, unpublished data).

Seven monoclonal antibody preparations specific for vp7 were studied in passive protection experiments (Table 1). The structural specificities of these monoclonal antibodies were demonstrated by immunoprecipitation, reactivity with virus reassortants or viral variants, or both (10, 26) (data not shown). Three of these monoclonal antibodies (159-H11, 4F8, and 4C3) neutralized serotype 3 rotaviruses RRV and SA11 by PRN and protected suckling mice against serotype 3 (RRV and SA11) but not against OSU or UK virus challenge. One monoclonal antibody (5H3) neutralized only the RRV serotype 3 strain by PRN and protected suckling mice against RRV but not against SA11, OSU, or UK virus challenge. Competition antibody binding assays and viral variant analysis indicated that monoclonal antibody 5H3 recognized an epitope distinct from that recognized by monoclonal antibodies 159-H11, 458, and 4C3 (Shaw and Greenberg, unpublished data). Monoclonal antibody 2C9 neutralized only Wa rotavirus by PRN and did not protect suckling mice against heterotypic rotavirus challenge. Monoclonal antibodies 60-F2 and 129-B8 were directed against an epitope on vp7 which is not involved in neutralization in vitro (11). This epitope is shared antigenically with most group A rotavirus strains (data not shown). Passive transfer of monoclonal antibodies 60-F2 or 129-B8 failed to protect newborns against challenge.

Monoclonal antibody 255/60, isolated from lymphocytes obtained after RRV immunization, precipitated the major inner capsid protein vp6. This monoclonal antibody has been found to react with the subgroup epitope of vp6 (10). Monoclonal antibody 255/60 did not neutralize rotaviruses by PRN or protect suckling mice against RRV challenge.

In the present study we demonstrated that rotavirus-specific monoclonal antibodies were capable of protecting newborn mice when inoculated orally 30 min before heterologous host rotavirus challenge. In addition, we found that monoclonal antibodies administered up to 2 h after challenge protected suckling mice against disease (data not shown). The viral amplification and viral shedding observed in mice infected with heterologous host strains are less than those observed in homologous host rotavirus infection (12, 21). Nevertheless, the epithelial cell histopathological changes in the small intestine and serotype-specific humoral immune response in mice infected with heterologous host rotaviruses are characteristic of rotavirus-induced gastroenteritis (21). In addition, the clinical disease induced by heterologous host rotaviruses in mice is indistinguishable from that observed in homologous host infection (12). Therefore, the molecular determinants of antibody-mediated protection observed in our studies are likely to be relevant to the determinants of protection against challenge in natural rotavirus infection. However, the degree to which our findings are predictive of the ability of passively transferred rotavirus-specific monoclonal antibodies to either ameliorate or prevent homologous host rotavirus infection awaits further study.

Protection against challenge mediated by two or more epitopes on a single protein (4, 8, 24) or by distinct epitopes on two proteins (2, 8, 24, 25, 32) has been found in orthomyxoviruses, paramyxoviruses, herpesviruses, and alphavirus infections. We found that monoclonal antibodies directed against two distinct rotavirus surface proteins were capable of protecting suckling mice against challenge. A vp3-specific monoclonal antibody passively protected against diarrhea induced by three distinct rotavirus...
serotypes (OSU, RRV, and UK). Previous investigators found that monoclonal antibodies specific for vp3 of human rotavirus neutralized several human rotavirus serotypes in vitro (29). The epitope or epitopes recognized by protection against rotavirus-induced diarrhea in vivo awaits further study. On the other hand, we found that monoclonal antibodies specific for two epitopes of vp7 of serotype 3 rotaviruses protected newborn mice only against challenge with serotype 3 strains. A comparison of the mechanisms by which antibodies directed against vp3 and vp7 neutralize rotaviruses in vitro and protect against challenge in vivo awaits further study.

In this initial study, a monoclonal antibody specific for vp3 was capable of protecting suckling mice against challenge with three rotavirus serotypes. If vp3 is serotypically less diverse than vp7, this protein may represent a more rational target for vaccine strategies which use cloned rotavirus genes (1, 5, 7, 9). Alternatively, immunization with a reasortant rotavirus containing vp3 and vp7 from two antigenically distinct rotavirus parents might protect against diarrhea induced by two or more rotavirus serotypes.

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LITERATURE CITED


